

Madrid, 9 de mayo de 2012

USCAP
& AACR
HIGHLIGHTS

*Avances en nuevos marcadores en cáncer
y en biología tumoral AACR 2012*

*Federico Rojo
IIS-Fundación Jiménez Díaz*



Guión de la presentación

- Avances en melanoma y BRAF
- Cáncer de mama: más allá de HER2
- NSCLC y EGFR
- Biología del cáncer: temas candentes

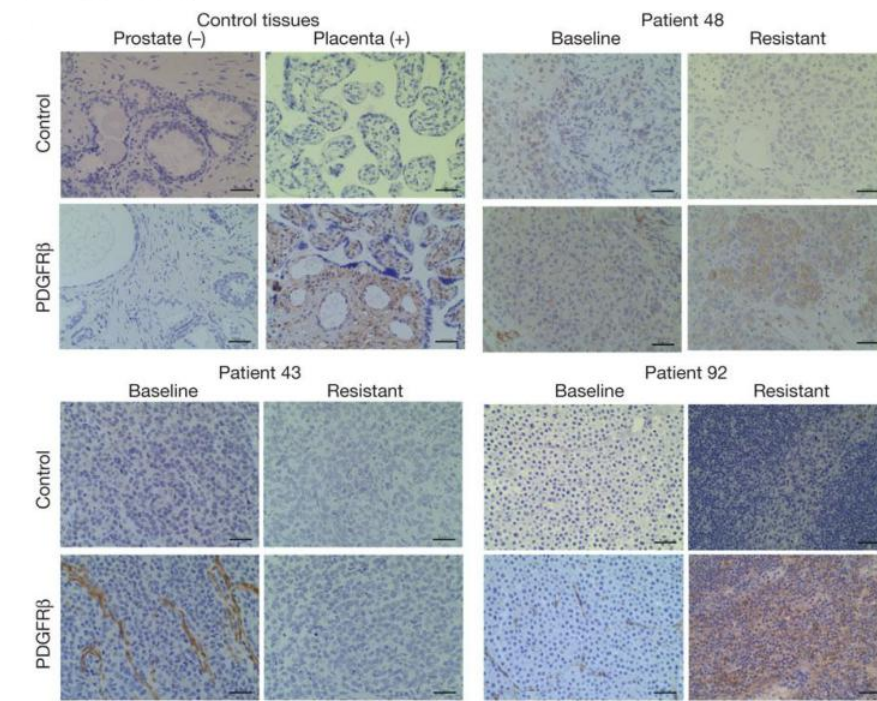
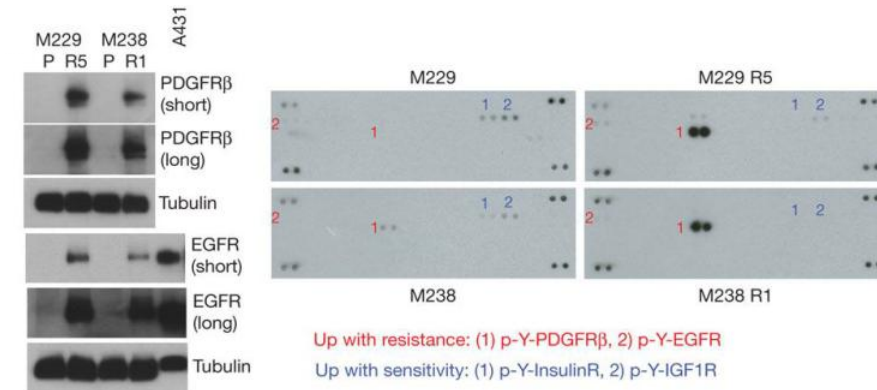
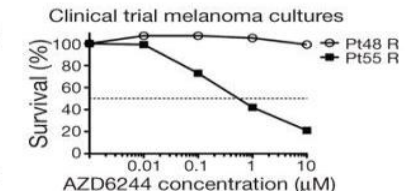
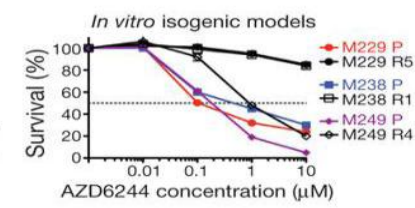
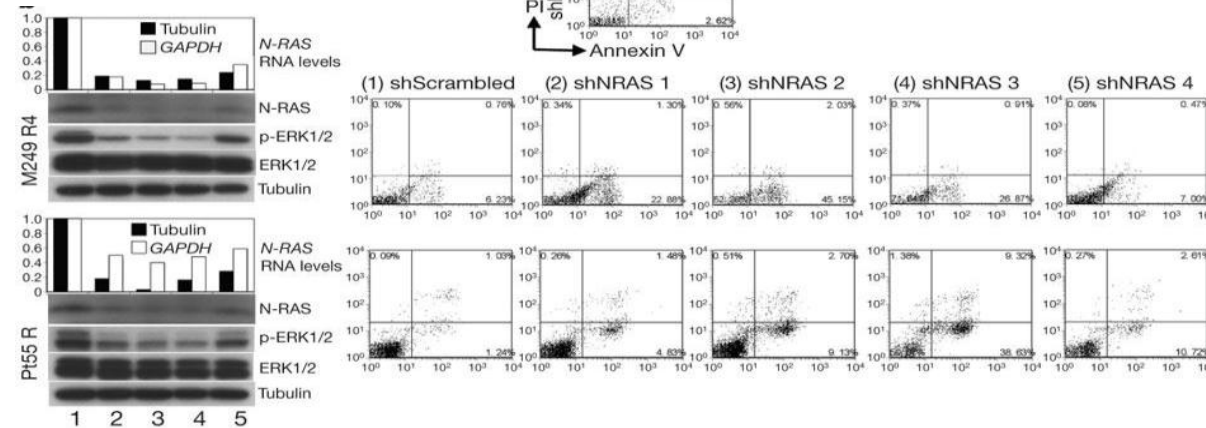
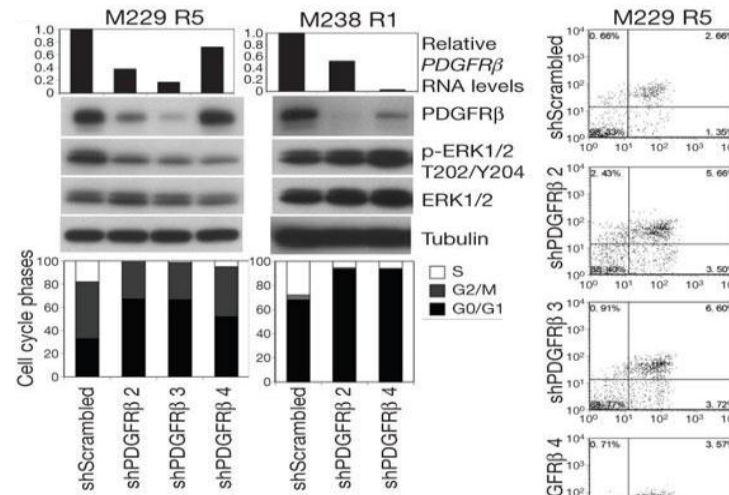
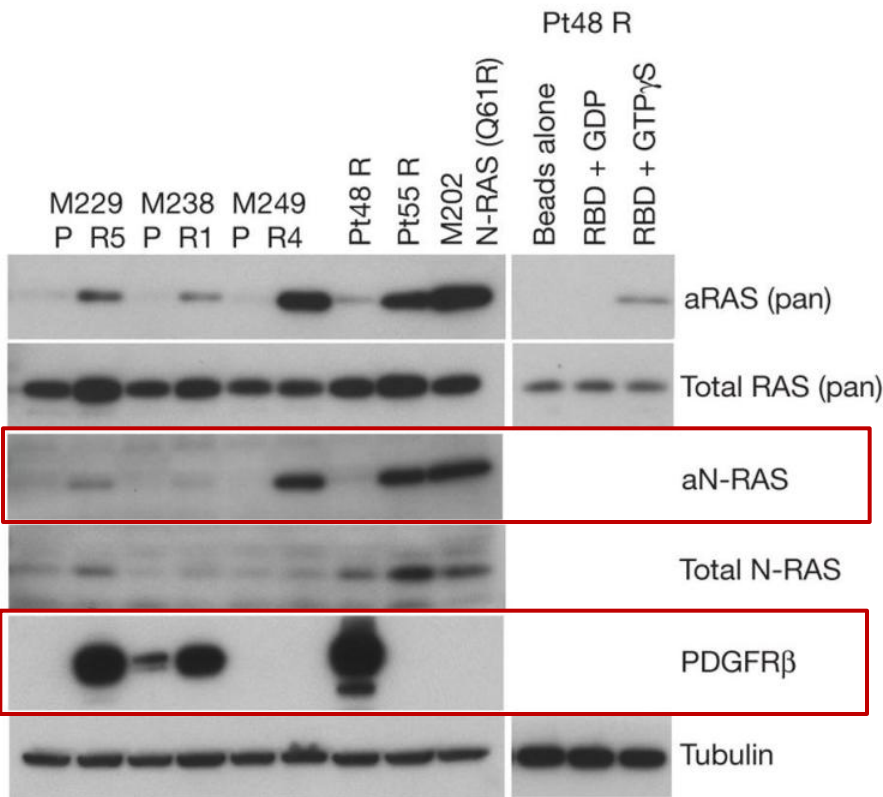
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BRAF biomarkers of sensitivity

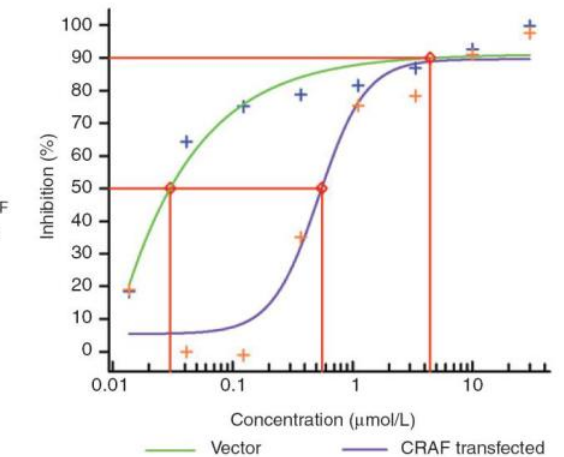
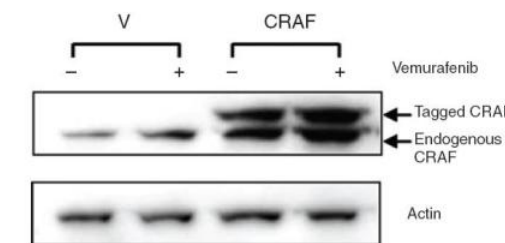
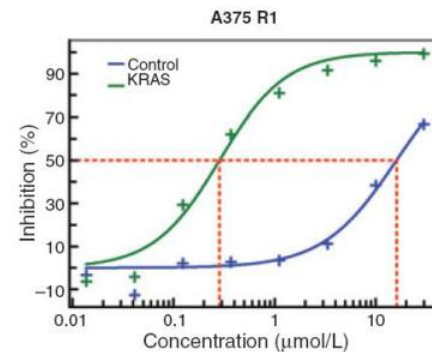
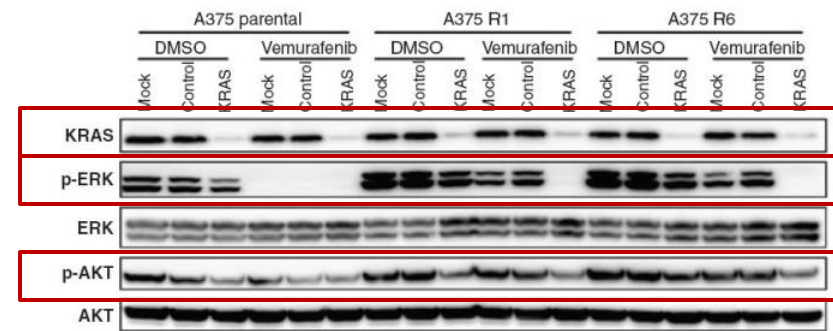
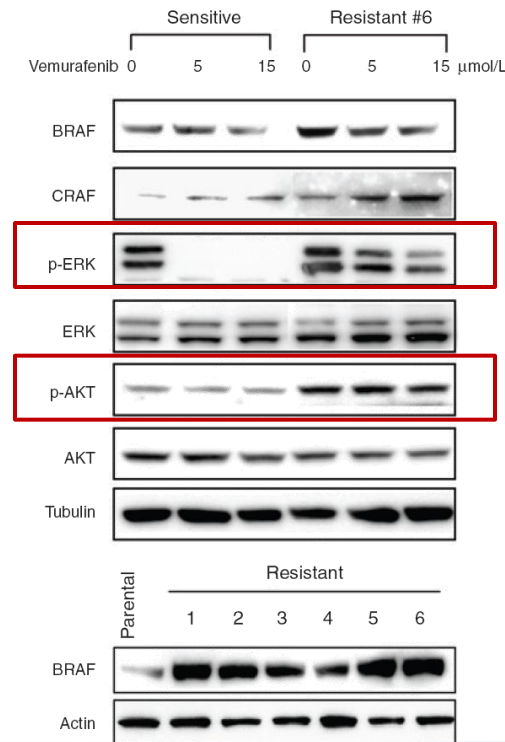
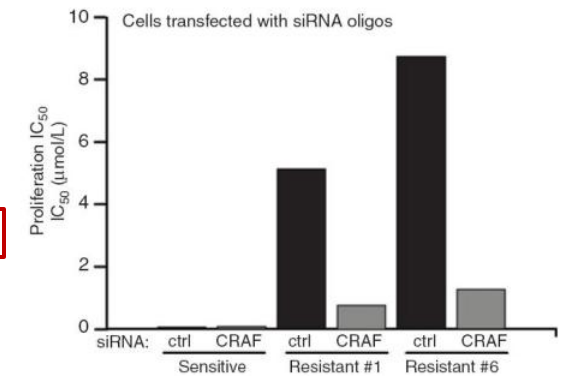
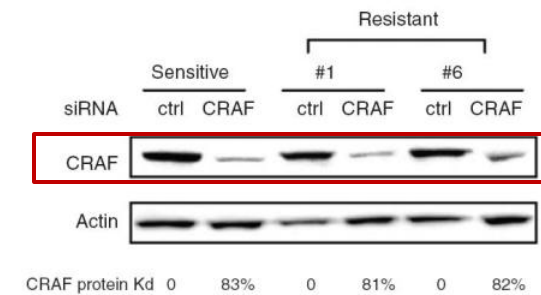
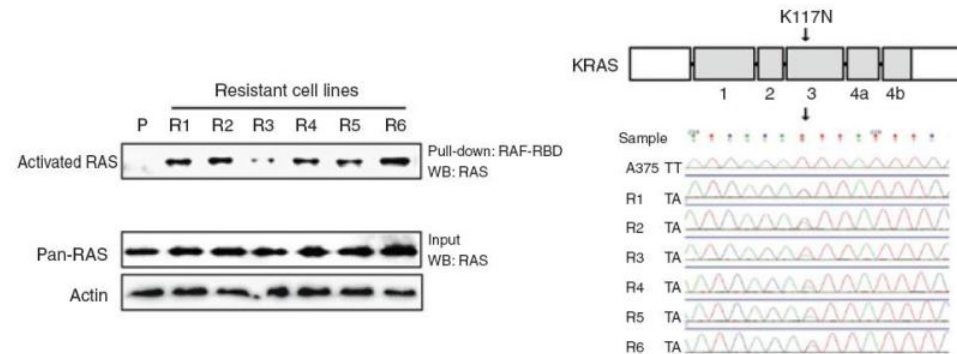
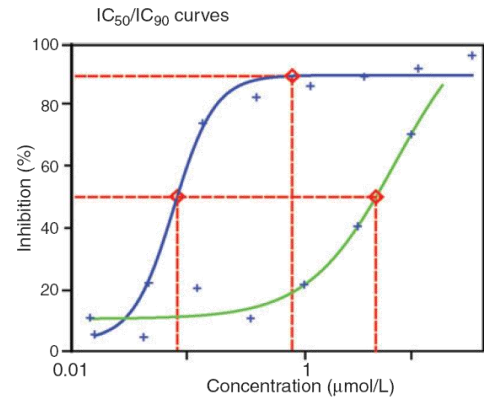
Targeting mutations in melanoma. Smalley, KSM

NRAS mutations in melanoma: How can we develop targeted therapies for these patients? Davies, MA



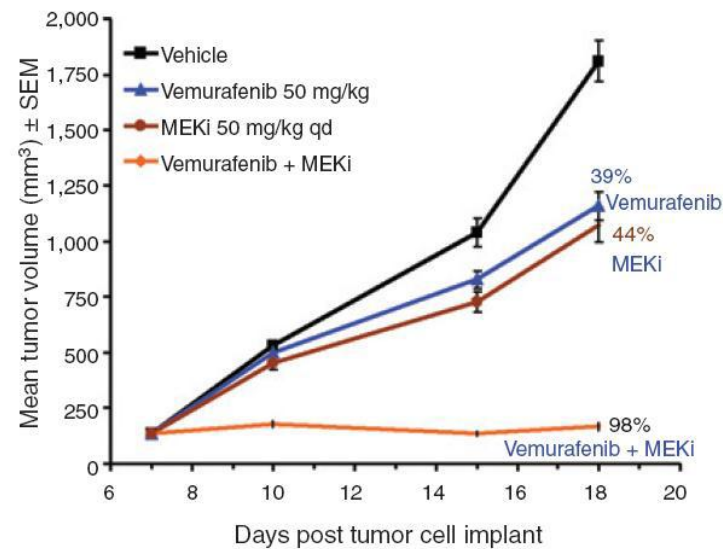
Resistance to Selective BRAF Inhibition Can Be Mediated by Modest Upstream Pathway Activation

#2806 BRAF and MEK inhibitor profiling across 240 tumor cell lines to correlate with sensitivity and resistant biomarkers. K Bernards, Y Ovechkina, J Cheng, J Kushleika, A Angione, J Hnilo, C O'Day.

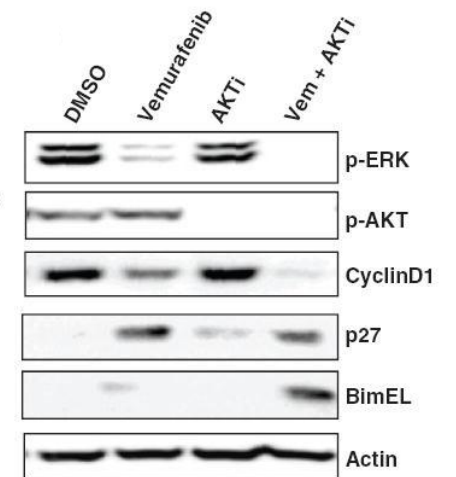
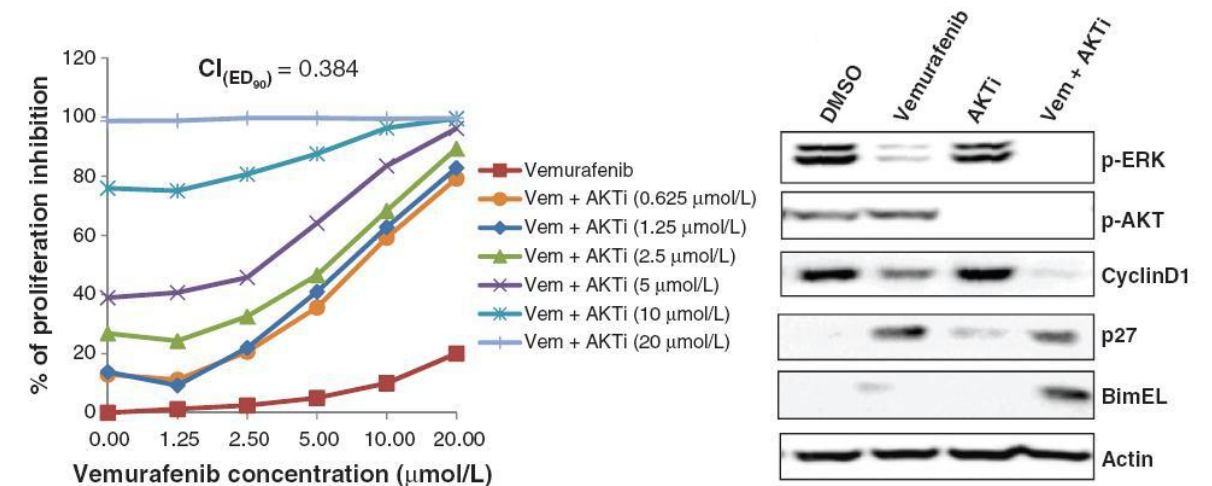
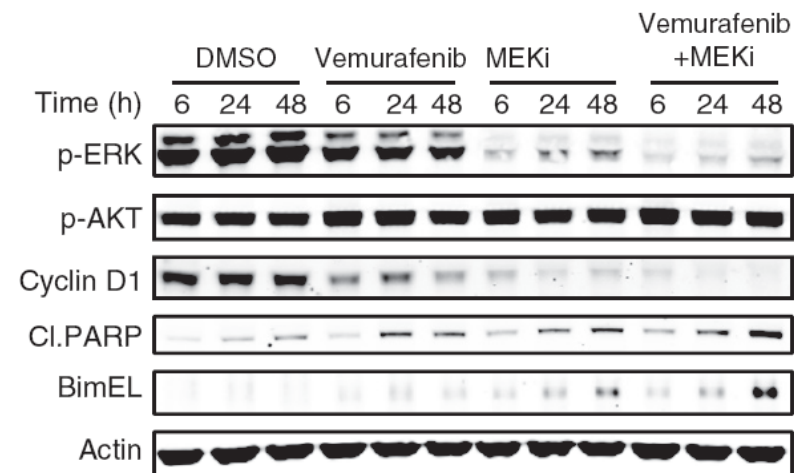


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Dose group (mg/kg, qd)	TGI (%)	PR	ILS (%)
Vehicle	-	-	-
Vemurafenib (50)	39	-	22
MEKi (12.5)	11	-	0
MEKi (25)	18	-	22
MEKi (50)	44	-	33
Vemurafenib (50) + MEKi (12.5)	88	1/10	78
Vemurafenib (50) + MEKi (25)	92	-	78
Vemurafenib (50) + MEKi (50)	98	3/10	100

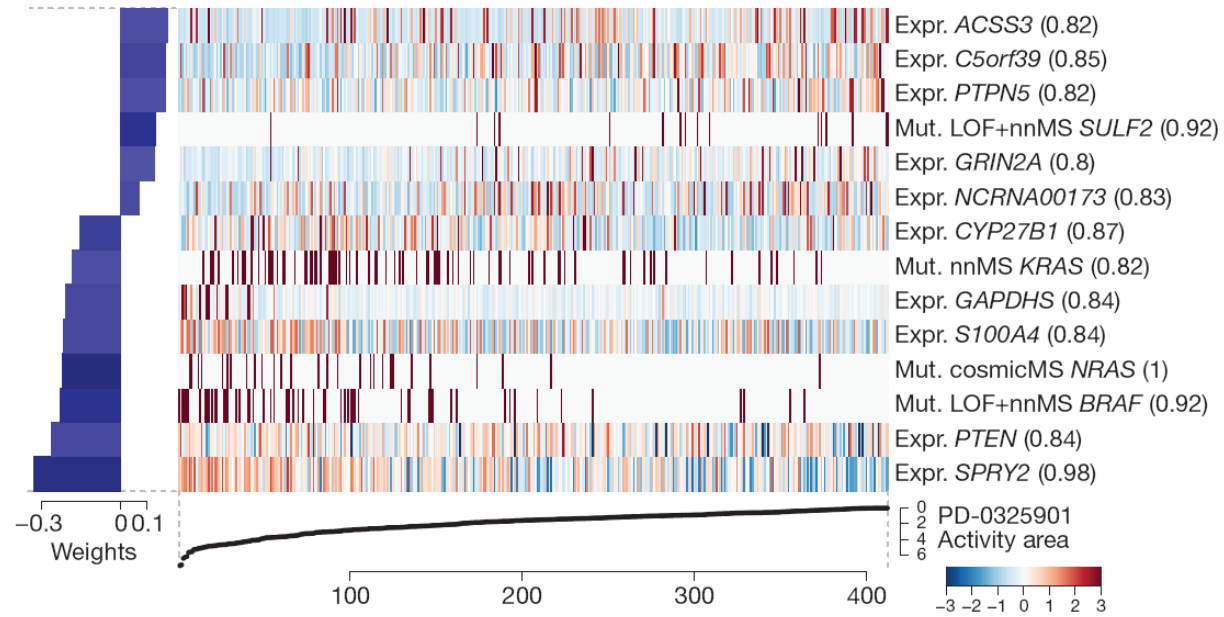
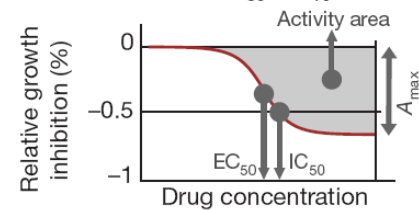
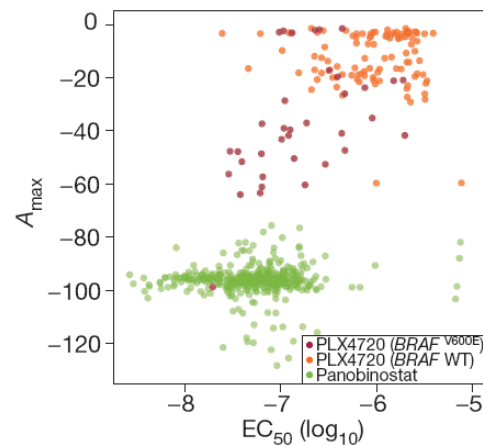
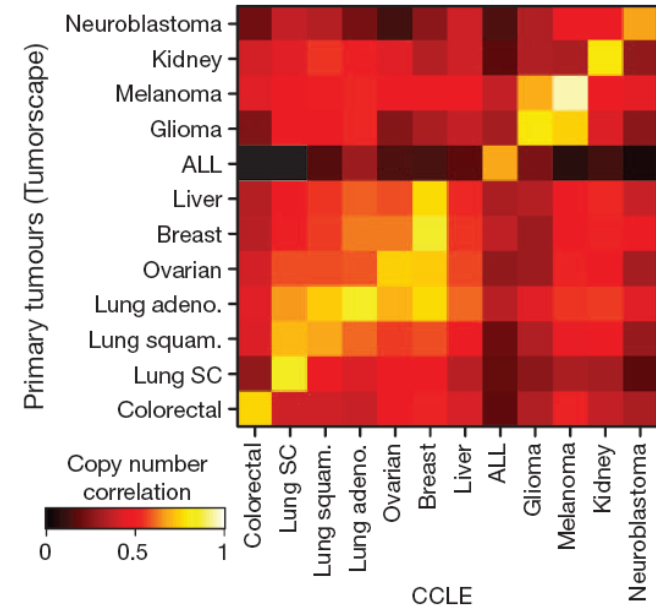
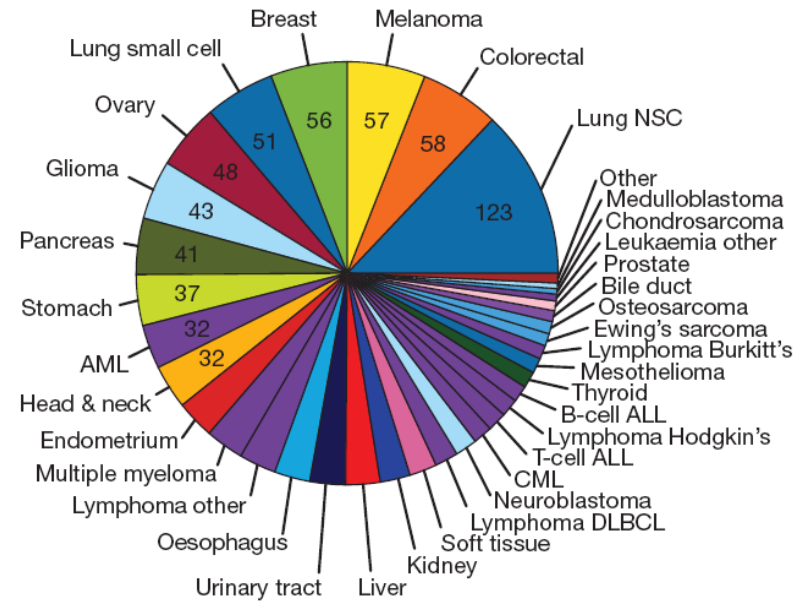


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- MEK1 mutations (MEKp124L/S, Emery, PNAS 2009; MEK1C121S, Wagle, JCO 2011)
- COT overexpression, CRAF activation (Johannessen, Nature 2010)
- NRAS mutation, PDGFR overexpression (Nazarian, Nature 2010)
- IGF1R overexpression (Villanueva, Cancer Cell 2010)
- BRAF amplification (Corcoran, Sci Signal 2010)

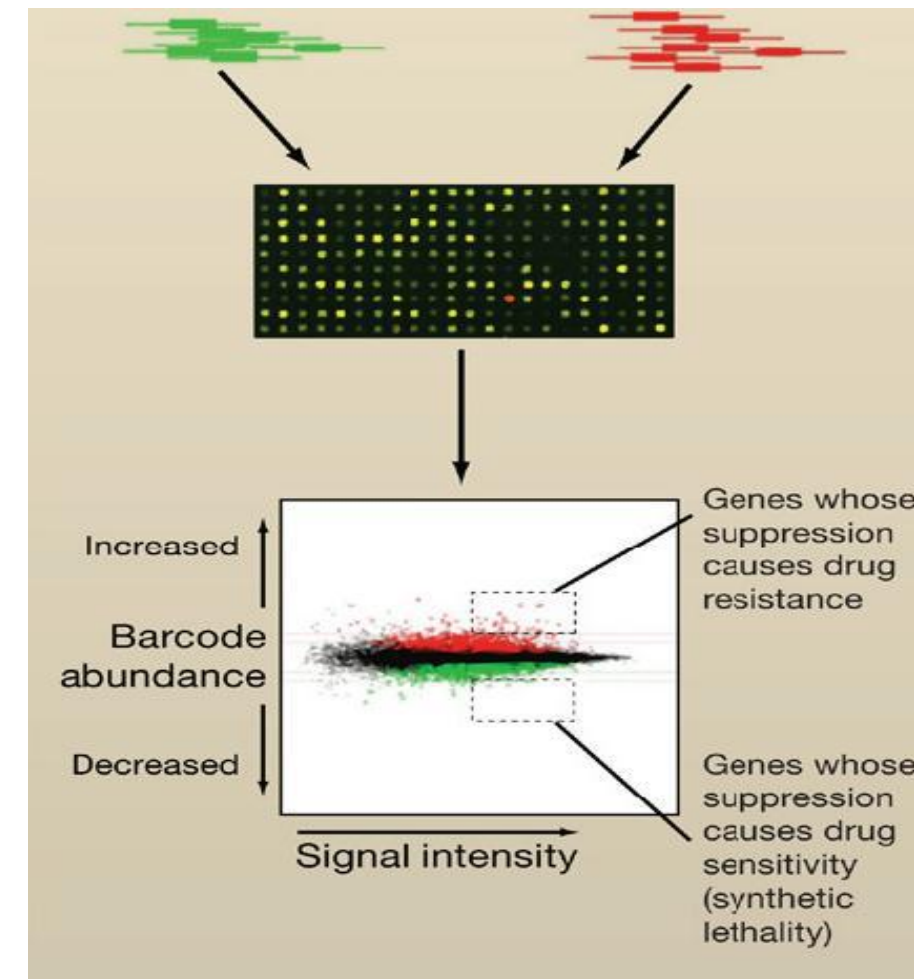
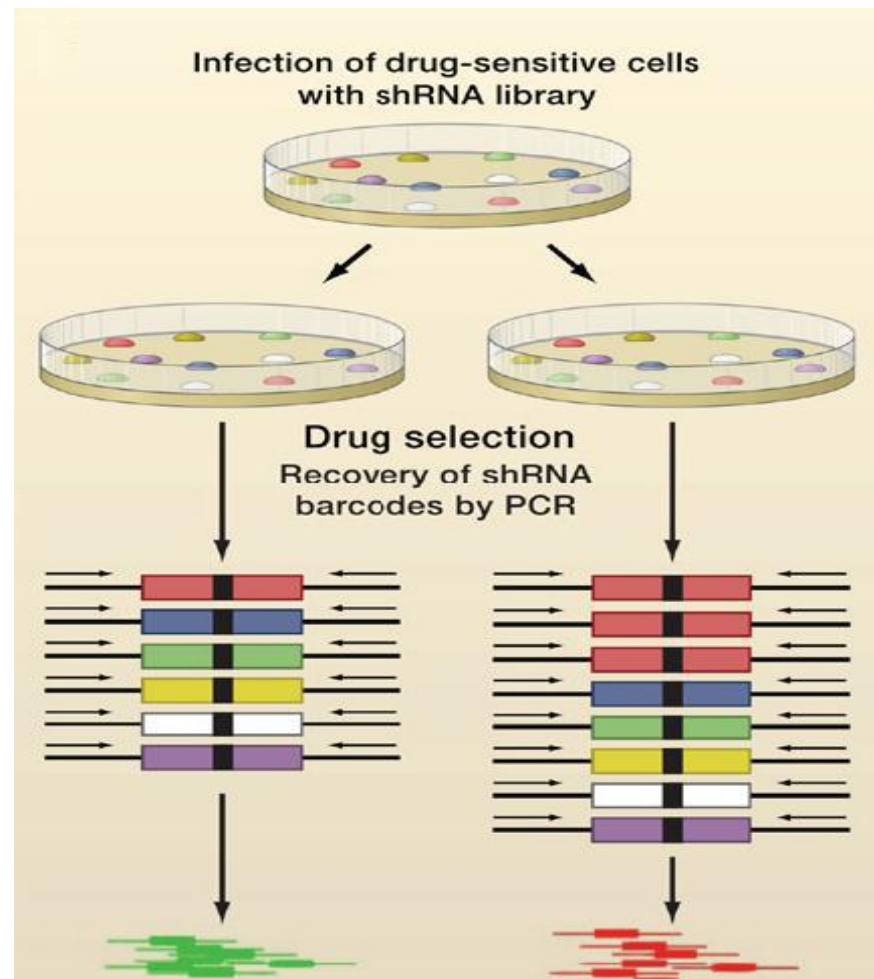
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Unresponsiveness of colon cancer to BRAF(V600E) inhibition through feedback activation of EGFR

#2156 Preclinical combinations of vemurafenib, a selective BRAF inhibitor, with other targeted therapies in BRAFV600E colorectal cancer models. H Yang, B Higgins, K Kolinsky, J Li, R Margolis, Z Go, A Railkar, K Packman, G Bollag, F Su.

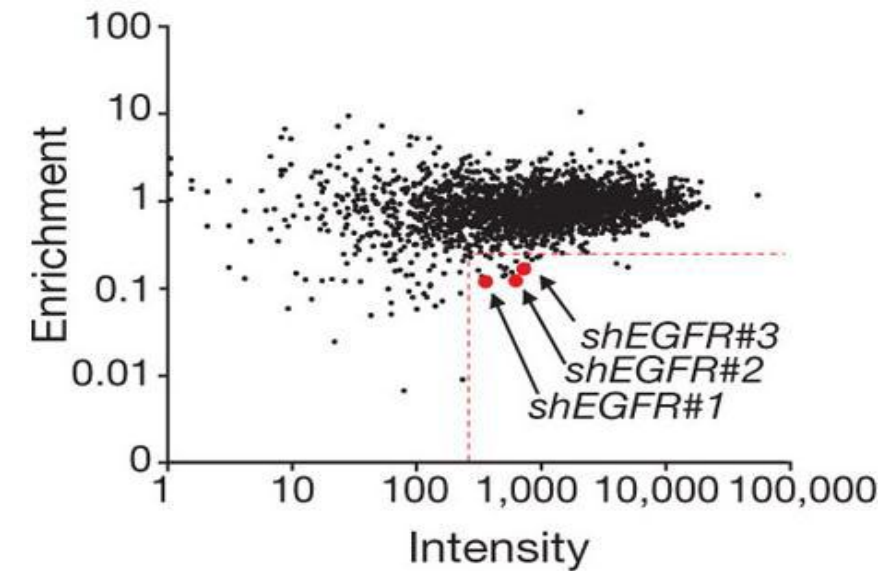
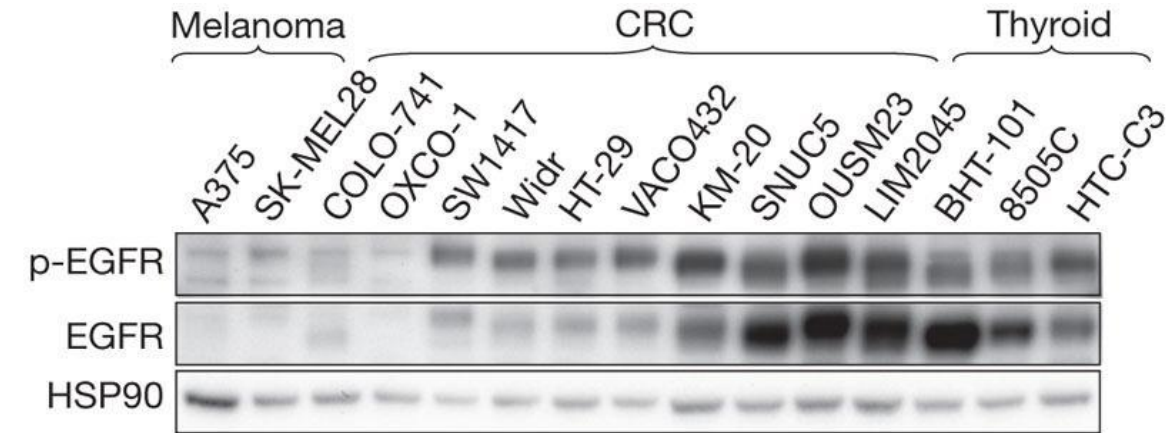
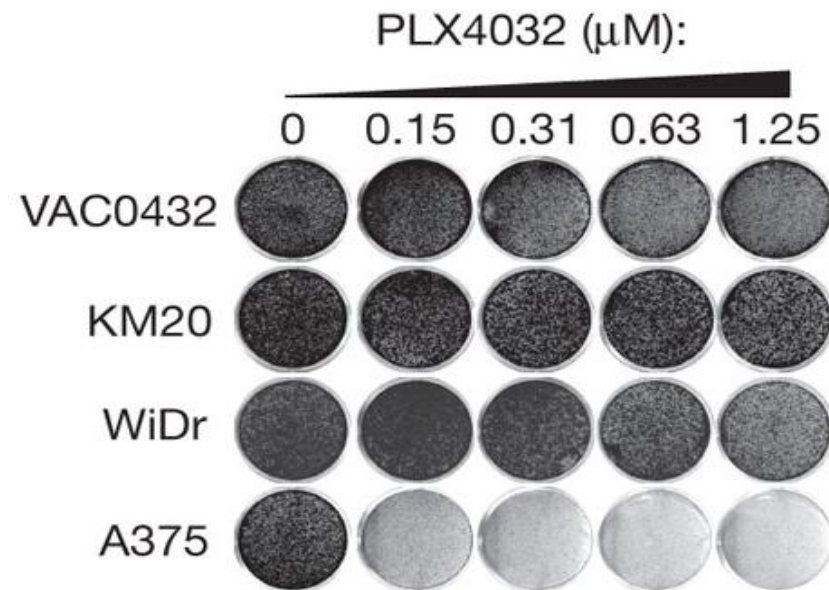
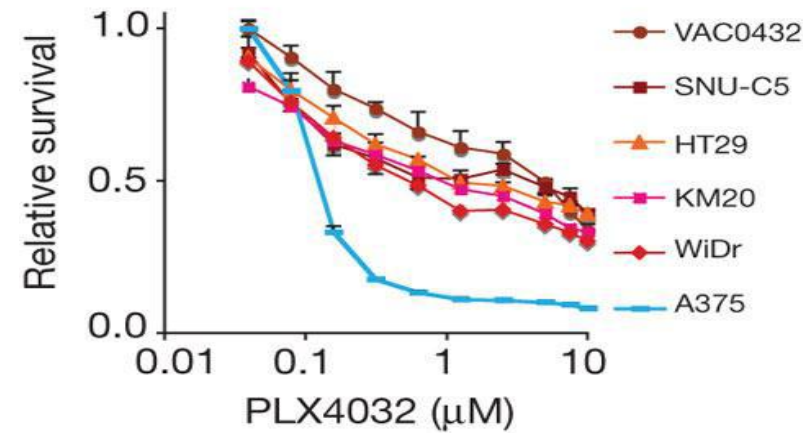
#5608 Functional genetic screens to find modulators of sensitivity to inhibitors of the PI3K/mTOR pathway. MS Van der Heijden, K Berns, MO Ports, R Bernards.



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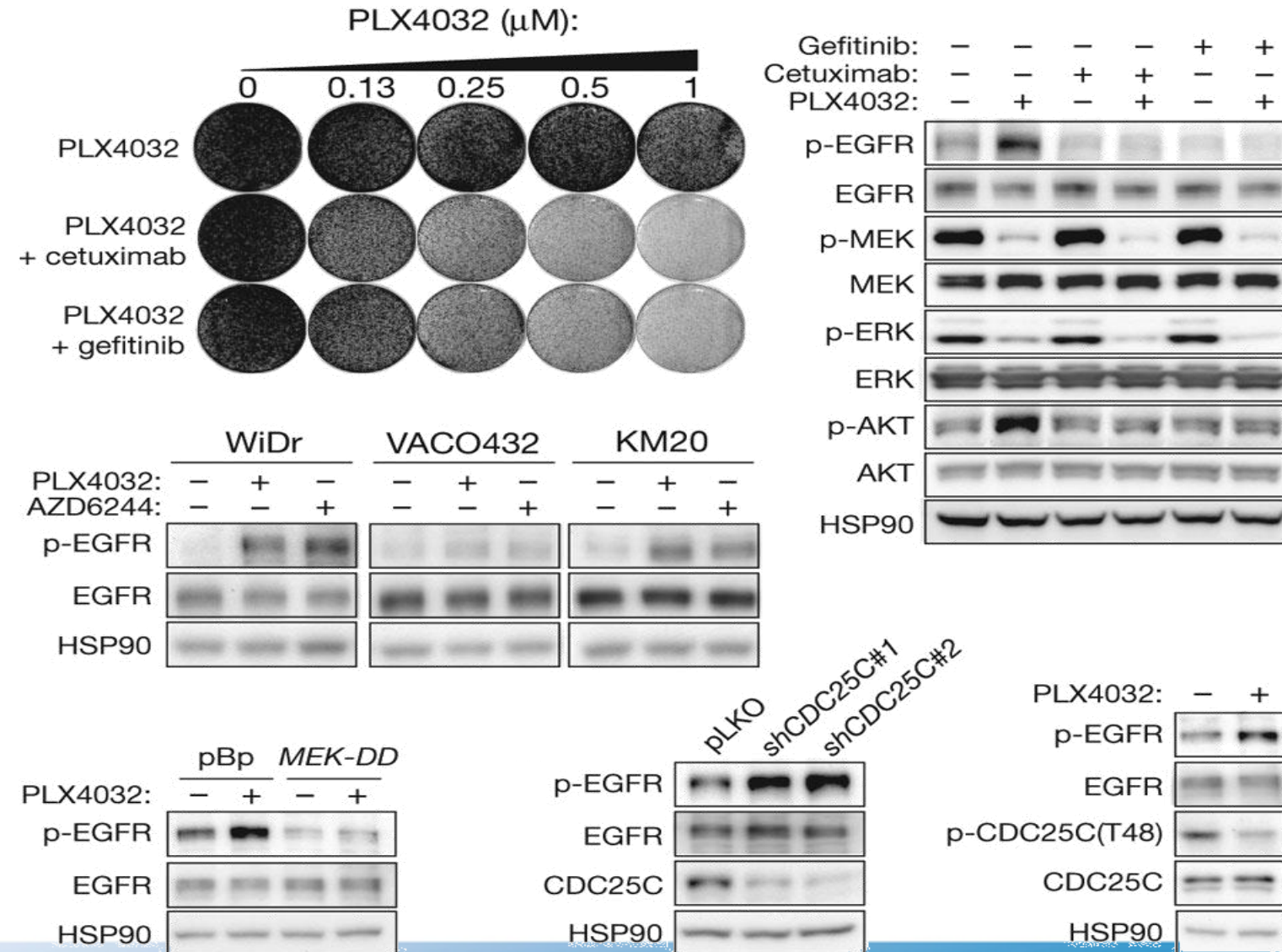
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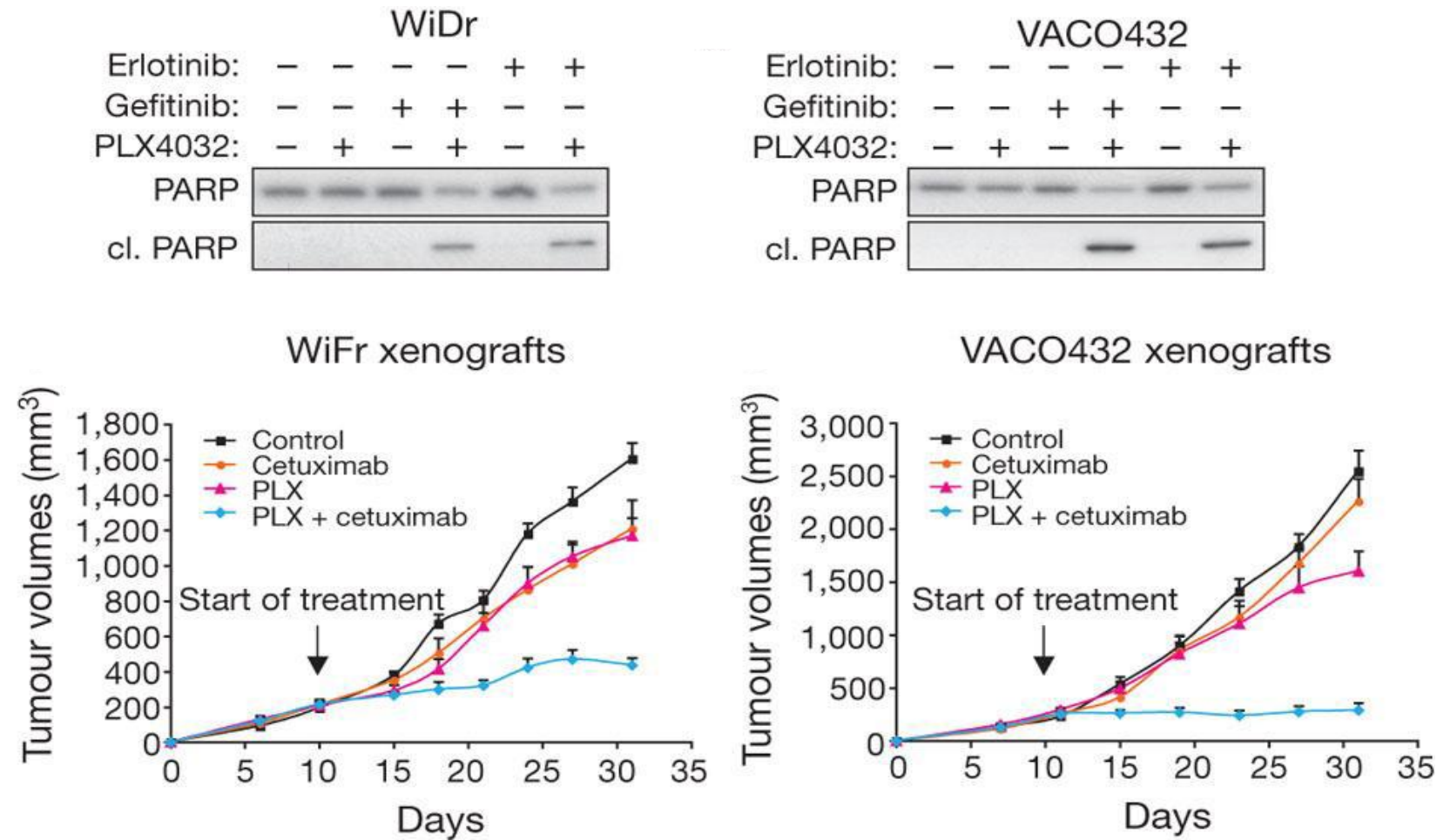
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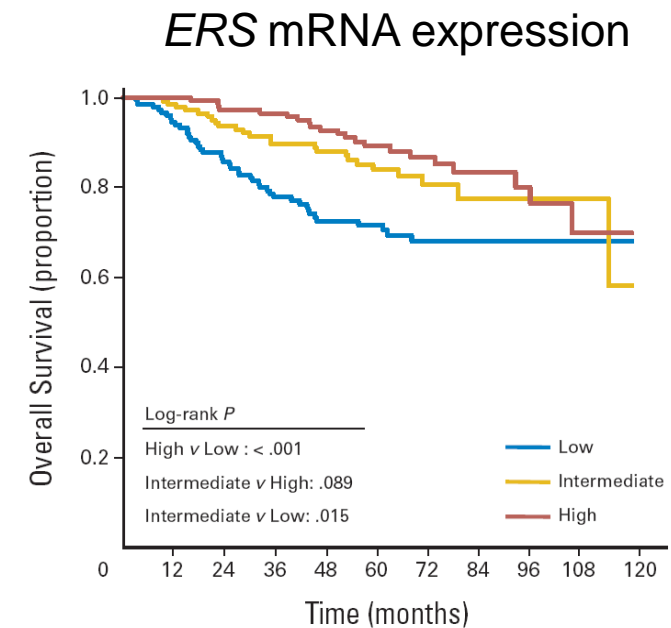
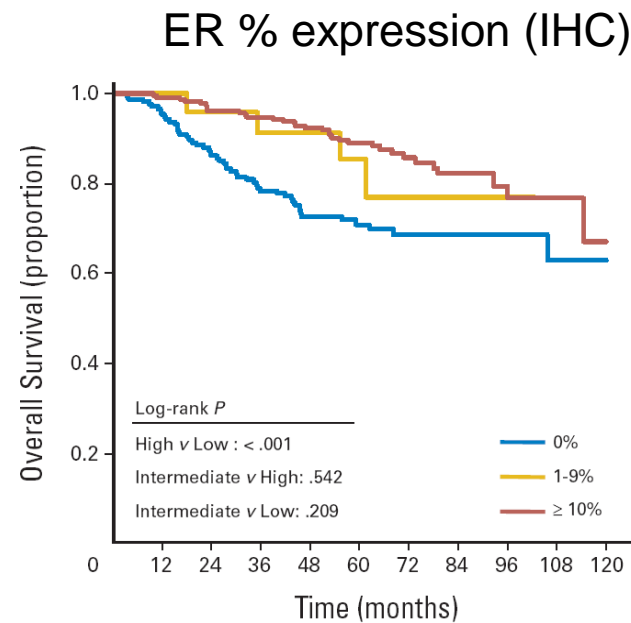
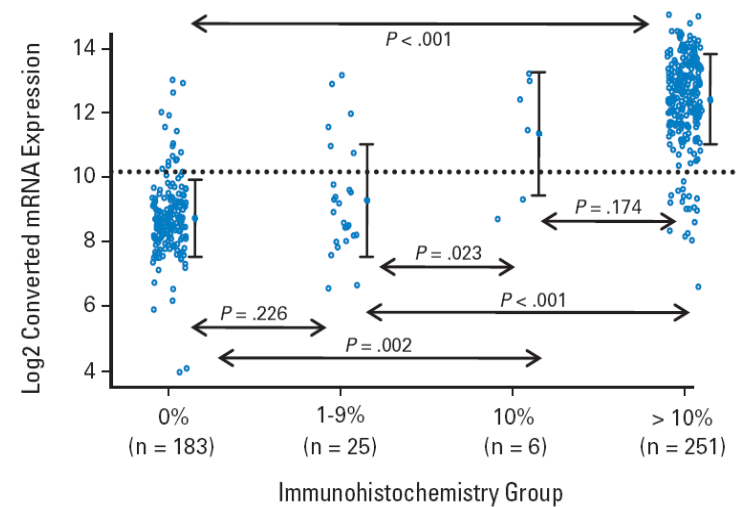
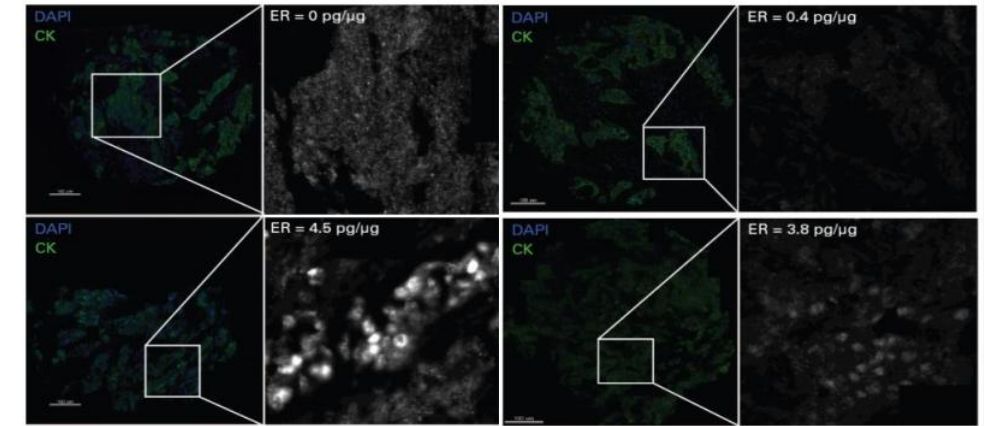
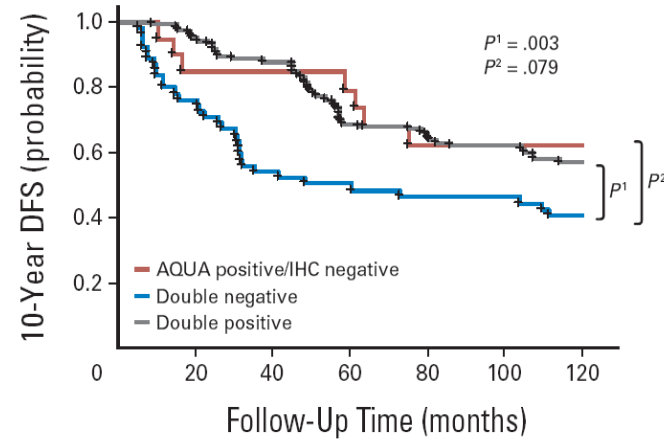
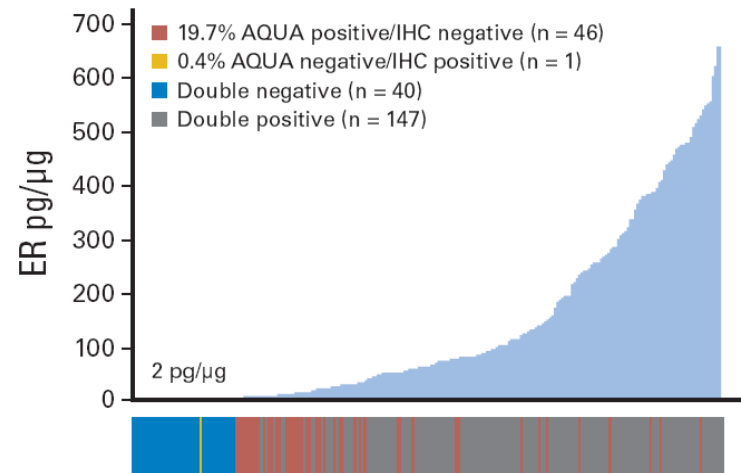


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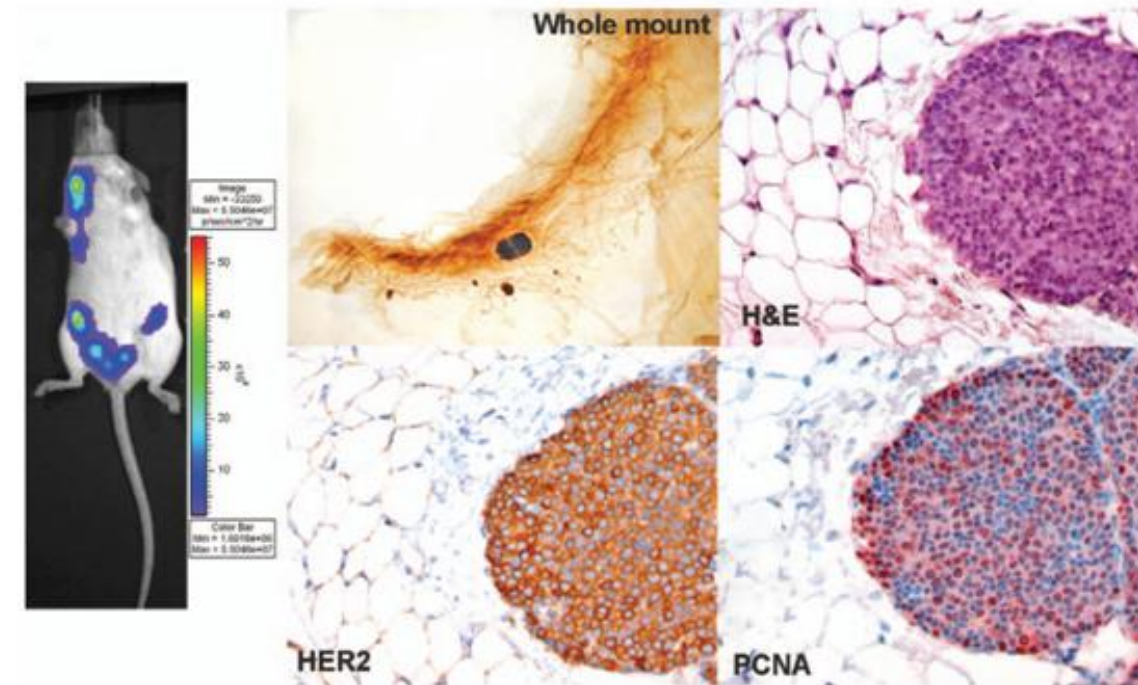
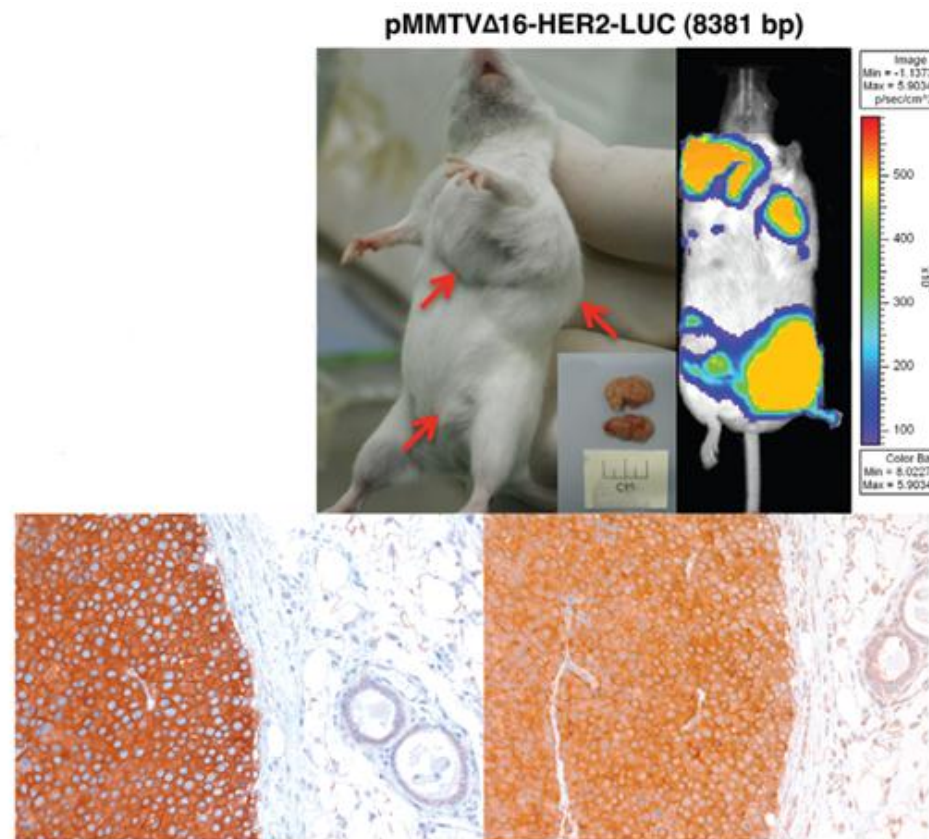
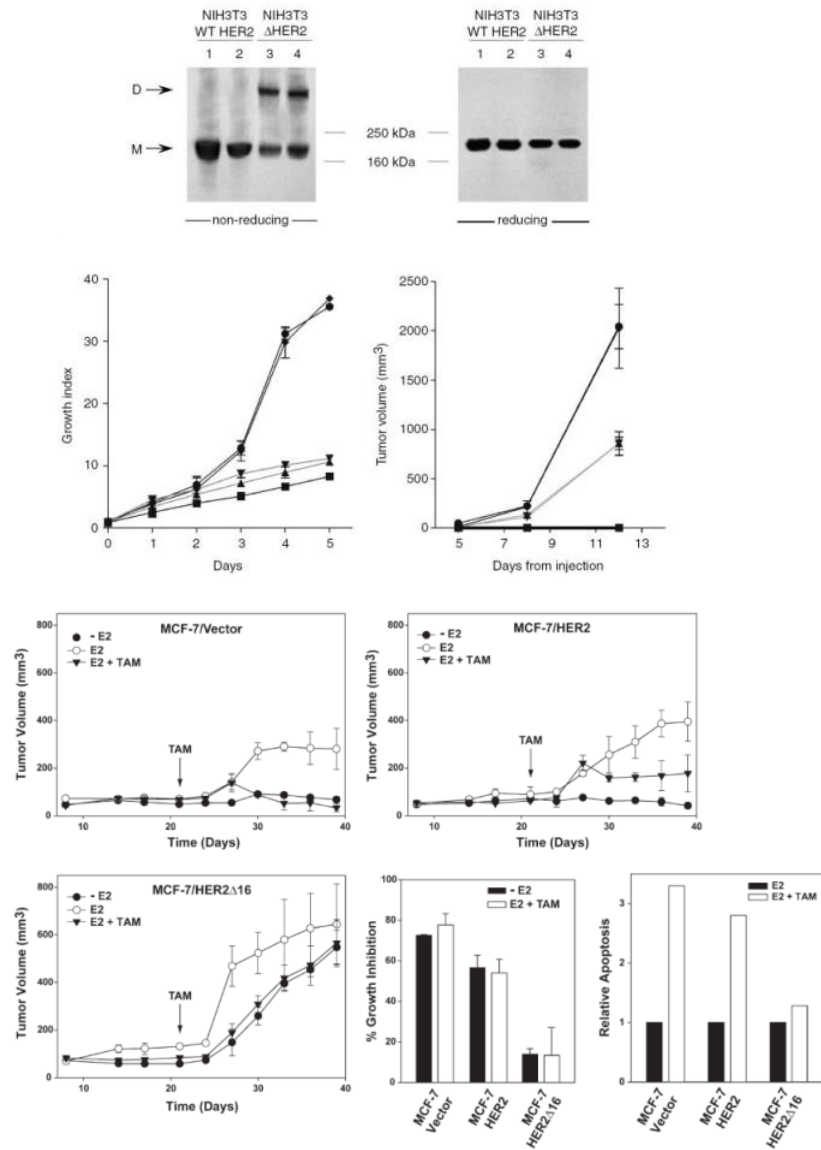
Estrogen receptor mRNA expression in breast cancer: correlation with response to tamoxifen

#691 Estrogen receptor mRNA level in breast cancer predicts response to tamoxifen. J Bordeaux, H Cheng, A Welsh, B Haffty, D Lannin, X Wu, N Su, X Ma, Y Luo, D Rimm



HER2 variants in breast cancer

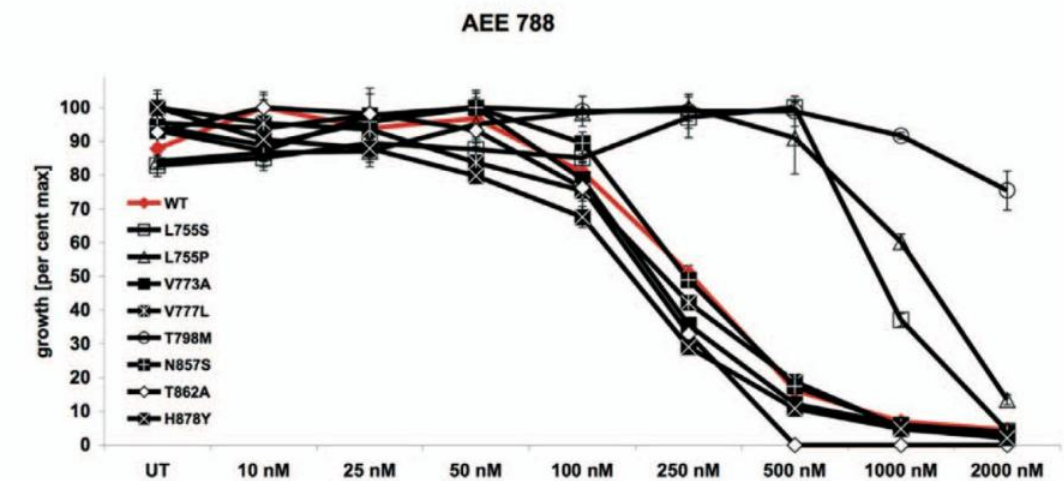
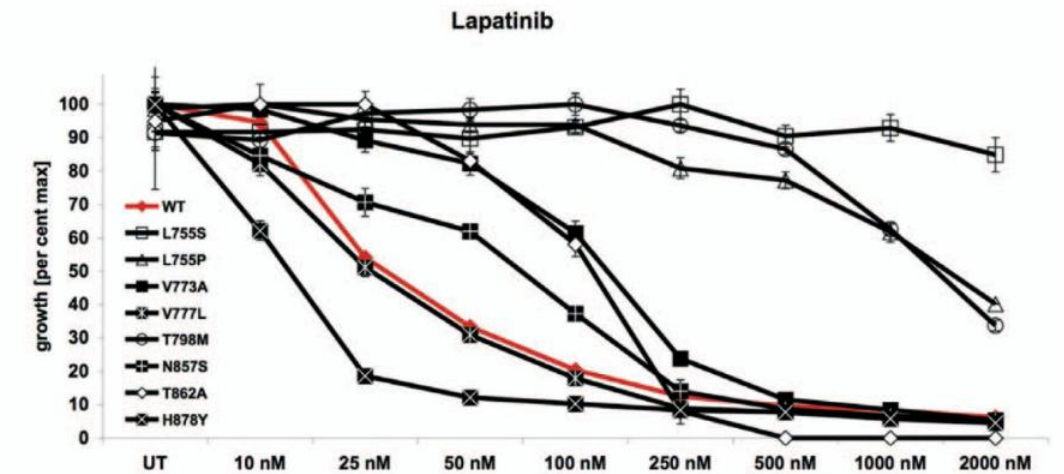
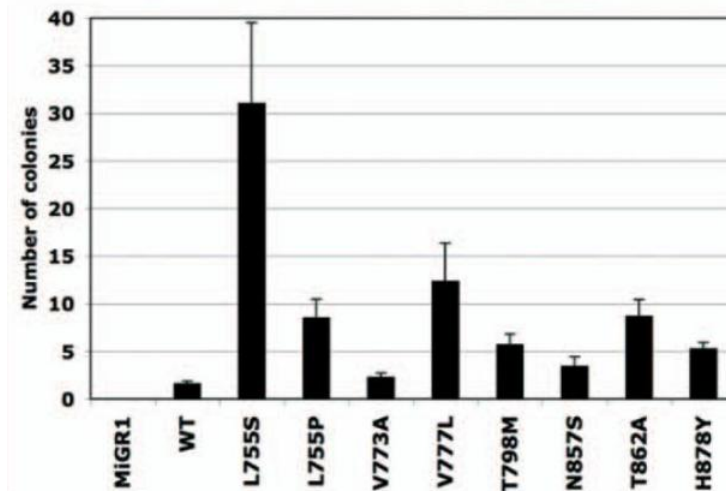
#916 Role of delta16HER2 splice variant in HER2-driven tumor progression and response to targeted therapy. SM Pupa, L Castagnoli, GC Ghedini, R Zappasodi, V Ciravolo, G Marzano, G Santilli, A Amici, C Marchini, M Di Nicola, R Canese, E Iorio, M Iezzi, E Tagliabue



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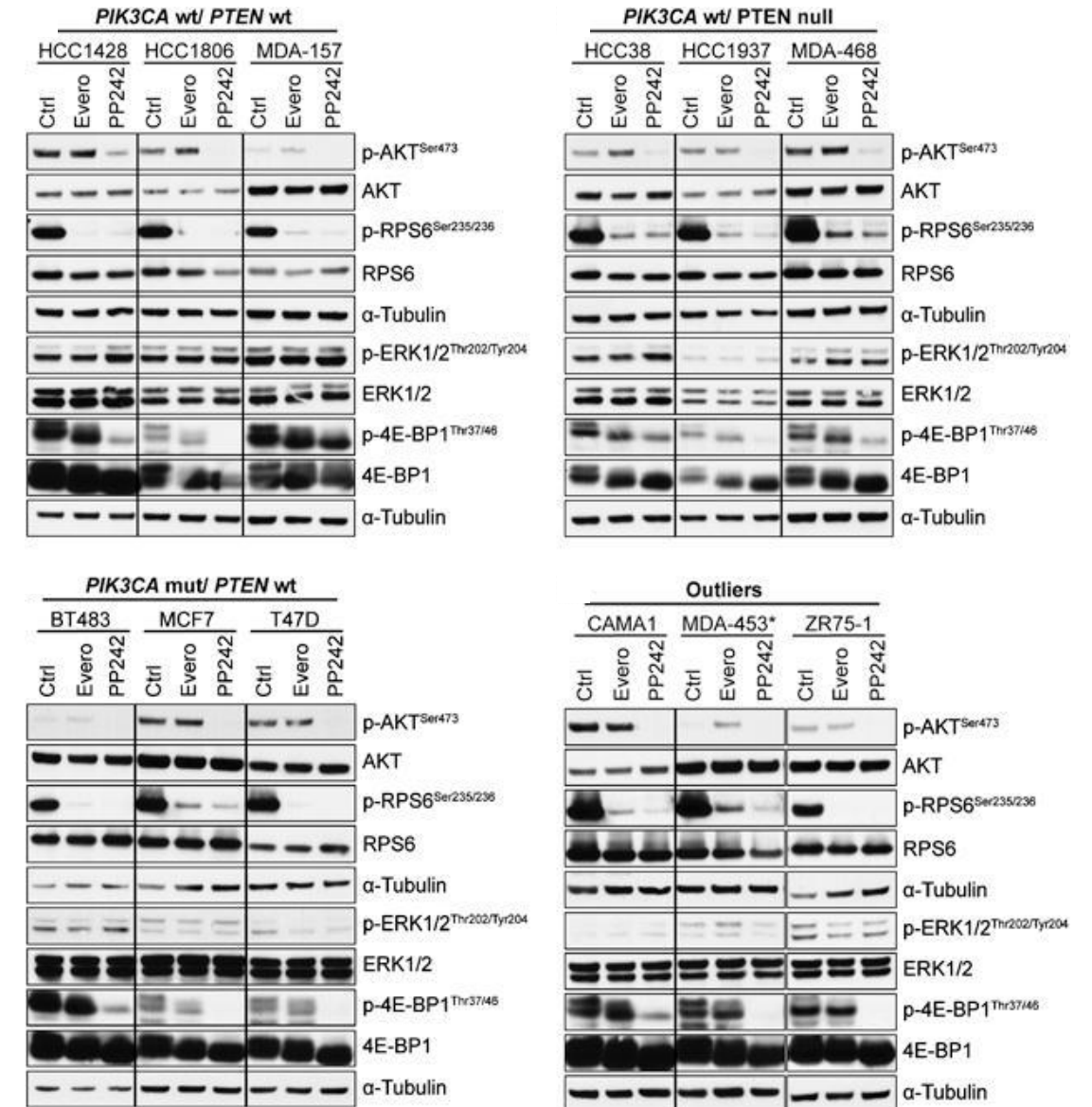
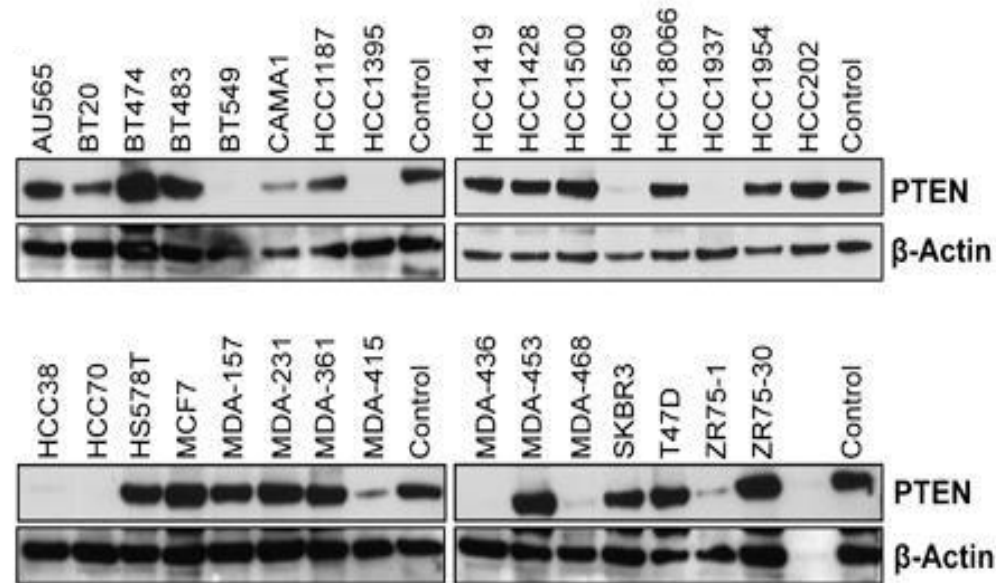
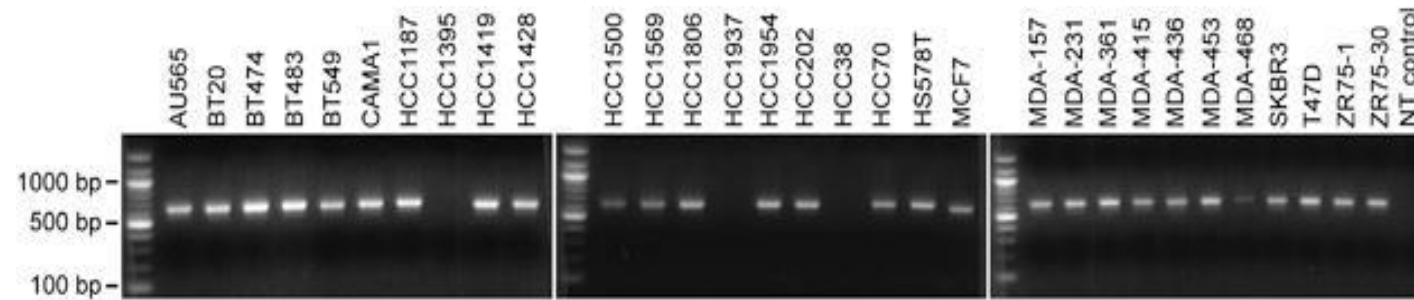
#3 *HER2 gene-amplified human breast cancer cells harboring a gatekeeper T768M mutation in HER2 overexpress EGFR ligands and are sensitive to dual therapeutic blockade of EGFR and HER2.* R Ghosh, A Narasanna, A Chakrabarty, JA Engelman, CL Arteaga

ERBB2 mutation	Exon	Functional region	Cancer type	Lapatinib	AEE788
WT	NA	NA	Breast cancer	30	257
L755S	19	ATP binding region	Breast and gastric cancer	>2000	897
L755P	19	ATP binding region	NSCLC	1545	1216
V773A	20	ATP binding region	SCCHN	146	200
V777L	20	ATP binding region	Gastric, colon and lung	27	215
T798M	20	Gate keeper residue	NA	1433	>2000
N857S	21	Activation loop	Ovarian cancer	75	246
T862A	21	Activation loop	Primary gastric cancer	125	191
H878Y	21	Activation loop	Hepatocellular carcinoma	14	168



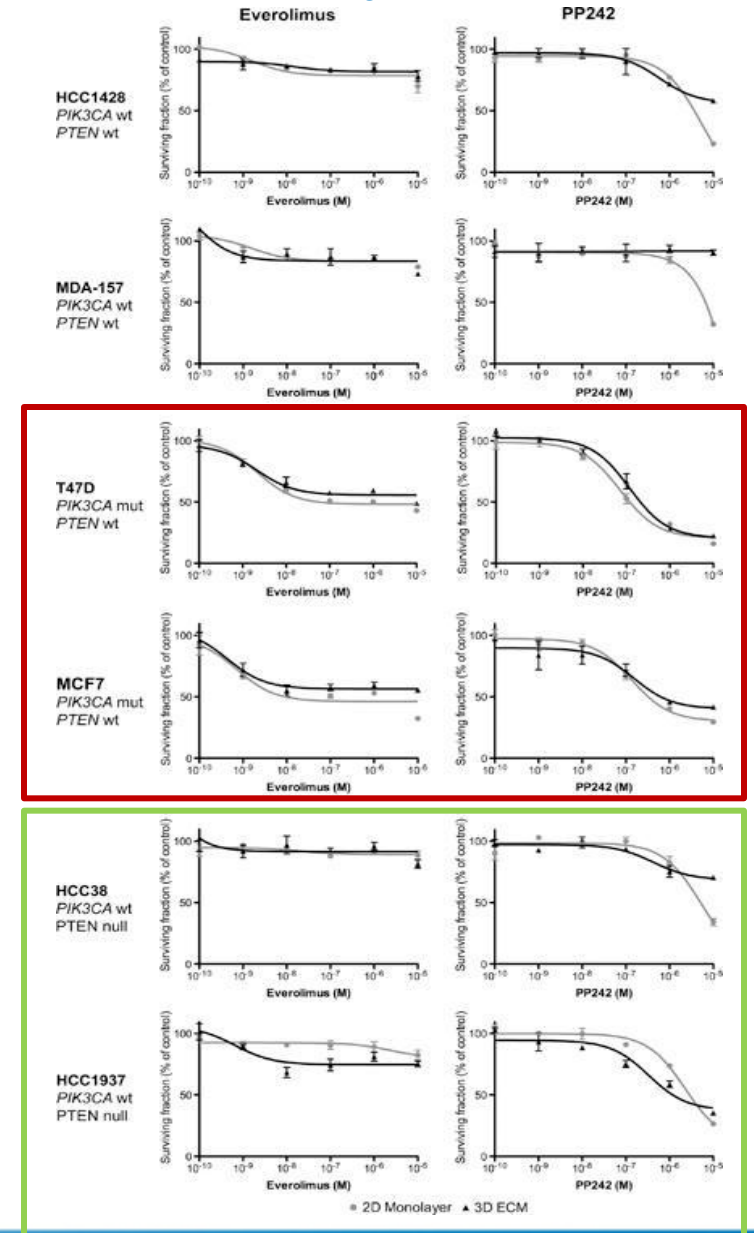
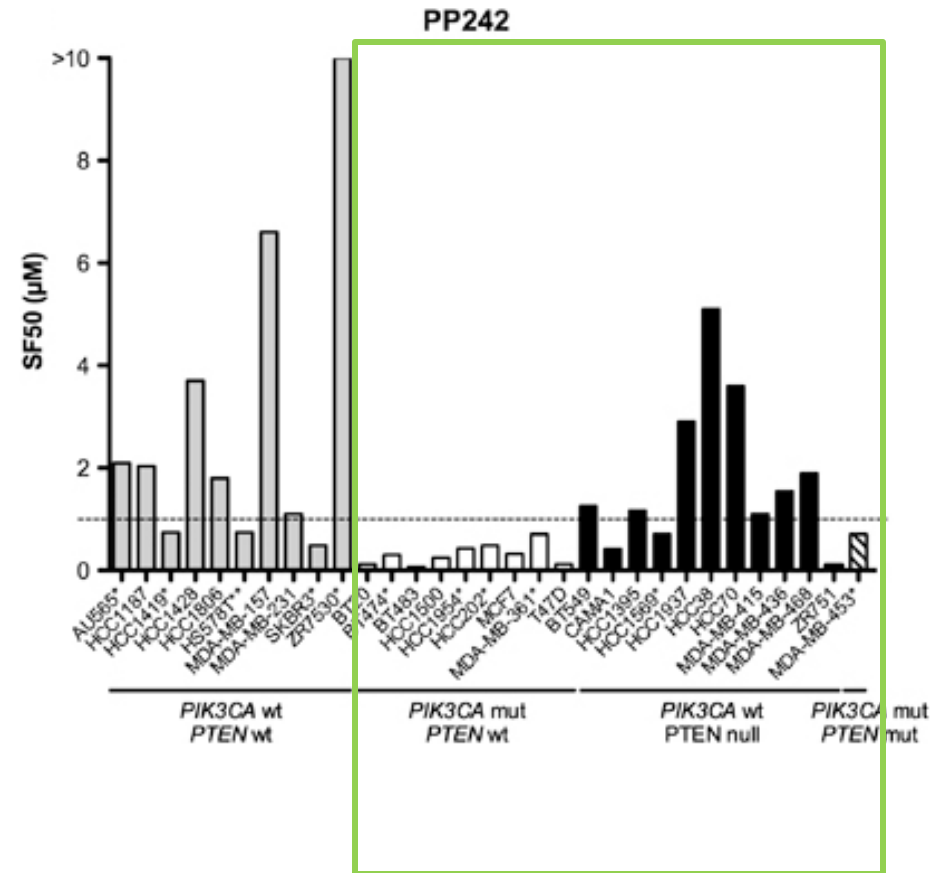
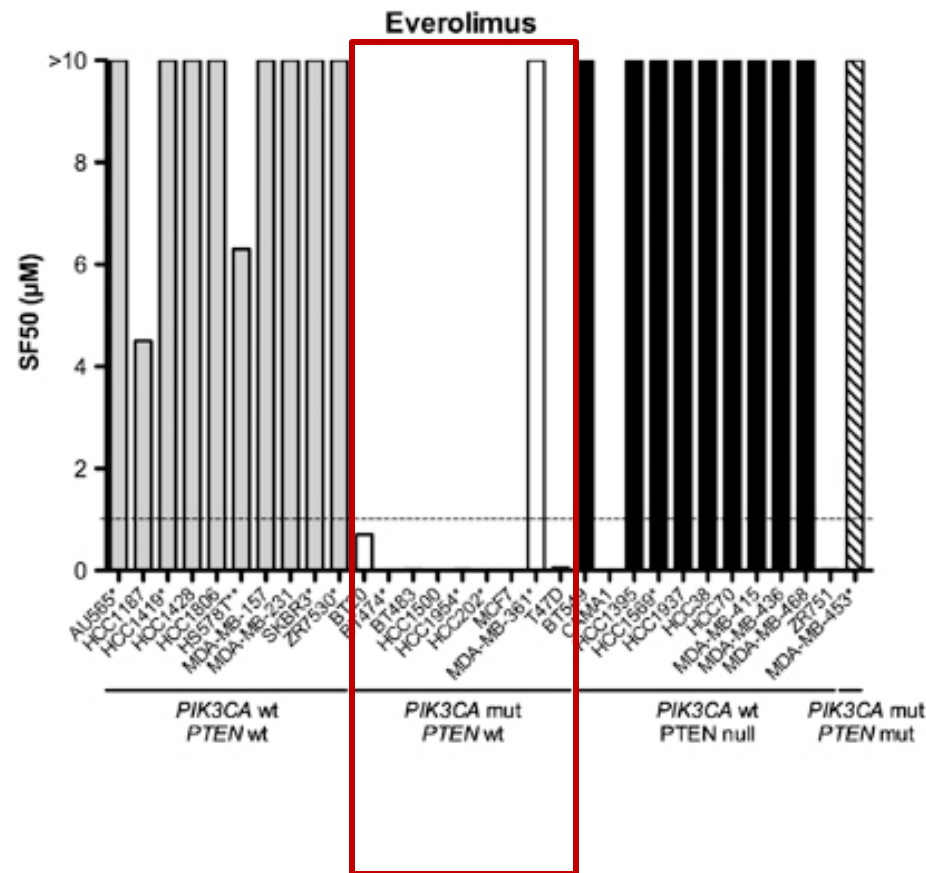
Predictive biomarkers for mTOR inhibitors

#4835 Genetic determinants of mTOR inhibitor response in breast and endometrial cancer. B Weigelt, PH Warne, M Lambros, JS Reis-Filho, J Downward



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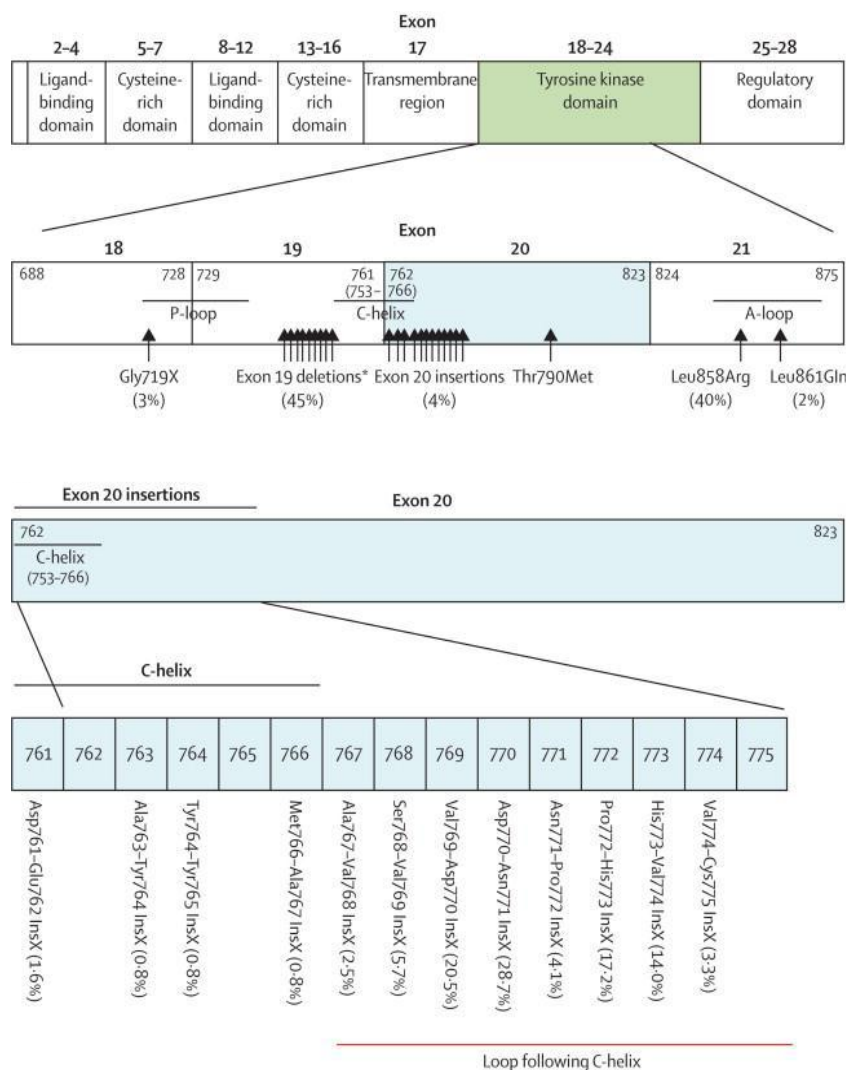


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Predictive role of EGFR exon 20 mutations in NSCLC

#23 Sensitivity of EGFR exon 20 insertion mutations to EGFR inhibitors is determined by their location within the tyrosine kinase domain of EGFR. H Yasuda, NJ Sng, WL Yeo, LL Figueiredo-Pontes, S Kobayashi, DB Costa.



EGFR mutation (number of mutations)	Number of patients (total=122)	Relative frequency (%)
762 Asp761_Glu762insGluAlaPheGln† (2)	2	1.6%
763 Ala763_Tyr764insPheGlnGluAla† (1)	1	0.8%
764 Tyr764-Val765insHisHis (1)	1	0.8%
765 -
766 Met766_Ala767insAlalle (1)	1	0.8%
767 Ala767_Val769dupAlaSerVal† (2), Ala767_Ser768insThrLeuAla (1)	3	2.5%
768 Ser768_Asp770dupSerValAsp§ (5), Ser768_Val769insValAlaSer (1), Ser768_Val769insAlaTrpThr (1)	7	5.7%
769 Val769_Asp770insAlaSerVal† (18), Val769_Asp770insGlyVal (1), Val769_Asp770insCysVal (1), Val769_Asp770insAspAsnVal (1), Val769_Asp770insGlySerVal (1), Val769_Asp770insGlyValVal (2), Val769_Asp770insMetAlaSerValAsp (1)	25	20.5%
770 Asp770_Asn771insSerValAsp§ (19), Asp770_Asn771insAsnProGly (1), Asp770_Asn771insAlaProTrp (1), Asp770_Asn771insAsp (1), Asp770_Asn771insAspGly (1), Asp770_Asn771insGly (5), Asp770_Asn771insGlyLeu (1), Asp770_Asn771insAsn (1), Asp770_Asn771insAsnProHis (1), Asp770_Asn771insSerValPro (1), Asp770_Asn771insSerValGln (1), Asp770_Asn771insMetAlaThrPro (1), delAsp770insGlyTyr (1)	35	28.7%
771 Asn771_Pro772insHis (1), Asn771_Pro772insAsn (1), Asn771_His773dupAsnProHis (1), delAsn771insGlyTyr (1), delAsn771insGlyPhe (1)	5	4.1%
772 Pro772_His773insProArg (11), Pro772_His773insTyrAsnPro (2), Pro772_His773insX (2), Pro772_His773insAspProHis (1), Pro772_His773insAspAsnPro (1), Pro772_His773insGlnVal (1), Pro772_His773insThrProHis (1), Pro772_His773insAsn (1), Pro772_His773insVal (1)	21	17.2%
773 His773_Val774insAsnProHis (6), His773_Val774insHis (3), His773_Val774insProHis (3), His773_Val774insGlyAsnProHis (2), His773_Val774dupHisVal (1), His773_Val774insGly (1), His773_Val774insGlyHis (1)	17	14%
774 Val774_Cys775insHisVal (4)	4	3.3%

	Best response to gefitinib or erlotinib		
	PR or prolonged SD	Total number of patients	% with PR or prolonged SD
Tyr764_Val765insHisHis	1 (patient had SD for 10 months)*	1	100%
Met766_Ala767insAlalle	1 (patient had SD for 10 months)†	1	100%
Ala767_Val769dupAlaSerVal	0	2	0%
Ser768_Asp770dupSerValAsp	1 (patient had PR for 24 months)‡	4	25%
Asp770_Asn771insAsp	0	1	0%
delAsn771insGlyTyr	0	1	0%
Pro772_His773insAsn	0	1	0%
Pro772_His773insVal	0	1	0%
Pro772_His773insTyrAsnPro§	0	1	0%
His773_Val774insGlyHis	0	1	0%
Exon 20 insertion (mutation type not reported)	0	4	0%
Exon 20 insertion (mutation type not reported)	0	1	0%
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Total	3 (1 PR, 2 prolonged SD)	20	15% (PR 5%)

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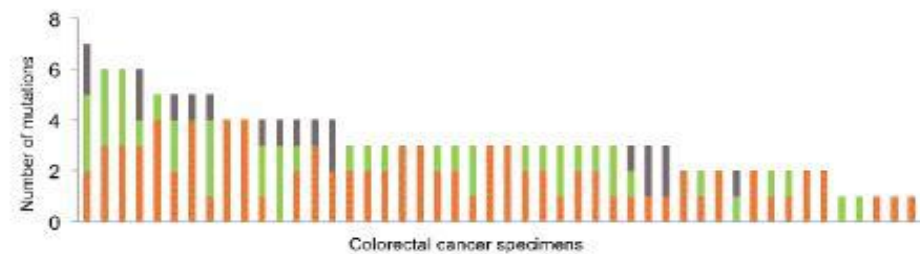
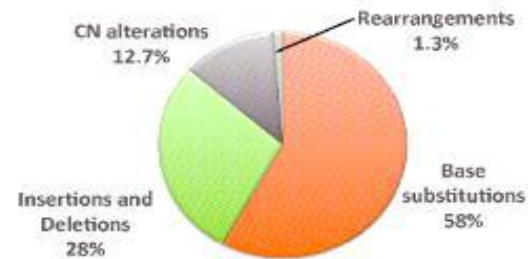
Comprehensive next-generation sequencing for clinically actionable mutations in FFPE cancer tissues

#3178 Can DNA from archived formalin-fixed paraffin embedded (FFPE) cancer tissues be used for somatic mutation analysis in next generation sequencing? E Palescandolo, R Jones, A Raza, A Sunkavalli, PK Brastianos, M Ducar, C Go, C Roden, C Hatton, M Hanna, A Mills, R Adusumilli, P Kumar, ML Meyerson, HC William, L MacConaill, P Van Hummelen

155 mutations found in 49 colorectal cancer specimens (of which 46 had previous KRAS testing)

KRAS testing in colorectal cancer: Concordance with reference lab

		FMI NGS Test	
		KRAS mutation positive	KRAS mutation negative
Reference Lab	KRAS mutation positive (N=46)	12	
	KRAS mutation negative		34



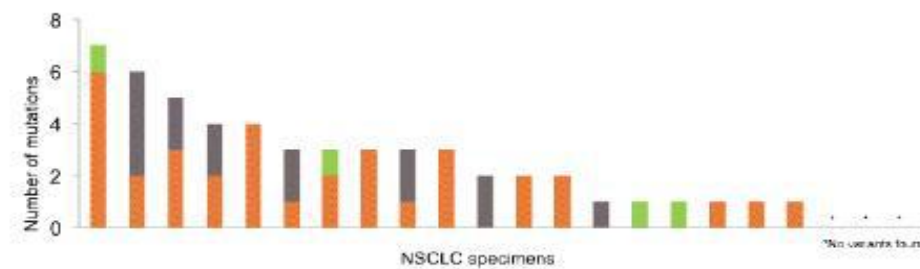
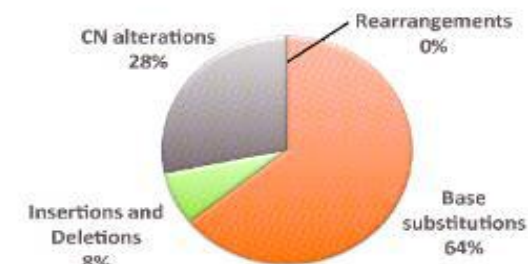
Conclusions

- This study confirms the ability to perform NGS with the small amounts of tumor tissue present in routine FFPE cancer specimens with sensitivity and specificity suitable for clinical decision-making. An average coverage of ~200X and 100% concordance was achieved with reference laboratories' conventional single gene analysis for *BRAF*, *KRAS*, and *EGFR*.
- Comprehensive cancer genomic testing identifies many more mutations than traditional methods. A total of 214 driver mutations were identified of which only 37 (17.6%) could have been found using conventional "hot spot" analysis.
- More than 50% of the specimens analyzed in this study contained mutations that could be used to inform cancer treatment decisions.

53 mutations found in 22 NSCLC specimens (of which 17 had previous EGFR testing)

EGFR testing in NSCLC: Concordance with 17 specimens tested by reference lab

		FMI NGS Test	
		EGFR mutation positive	EGFR mutation negative
Reference Lab	EGFR mutation positive (N=17)	1*	
	EGFR mutation negative		16



Variant Summary 75 Samples

Tumor Type	# specimens	Somatic mutations (avg)	Other variants (avg)	Comprehensive molecular analysis identified numerous known and novel mutations
Colorectal	49	3.2	7	TP53 (38), APC (32), KRAS (13), BRAF (6), SMAD4 (5), FBXW7 (4), PIK3CA (4), CDH1 (3), STK11 (3), BRCA1 (2), BRCA2 (2), ERBB2 (2), GNAS (2), ATM2 (1), CCR6 (1), FANCD1 (1), MAP2K4 (1)
NSCLC	22	2.4	8	KRAS (10), TP53 (6), STK11 (3), BRAF (2), MDM2 (2), PIK3CA (2), ATM (1), CDK4 (1), CDKN2A (1), CTNMB1 (1), EGFR (1), NF1 (1)
Melanoma	4	1.5	4	BRAF (2), TFS3 (2), CDKN2A (1)
All Types	75	2.85	7.1	

Data were obtained for a total of 75 patient specimens. An average of 2.9 known somatic mutations or likely somatic mutations were seen with a low of 1.5 in the melanoma specimens to a high of 3.2 in colorectal specimens. The number of other likely somatic variants detected averaged 7 across all specimens with a low of 4 in the melanoma and a high of 8 in the NSCLC specimens.

Intratumor heterogeneity and impact on biomarker detection

#5543 *Hetmap: Evaluating tumor heterogeneity in immunohistochemistry-stained breast cancer tissue.* SJ Potts, N Landis, D Eberhard, SC Schmechel, D Young, JS Krueger, H Lange

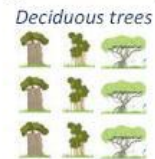
Consider a forest:



Each species of tree has a proportional abundance in a sample of that forest



A forest may be high in diversity



However, there is an important difference between *the types of trees* within a forest

Or a forest may be high in evenness

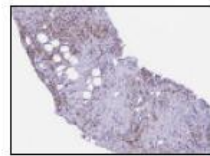


However, there is an important difference between *the types of cells* within a tissue

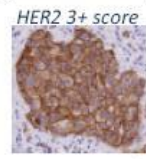
Consider a breast biopsy:



Each type of cancer cell has a proportional abundance in a sample of that tissue section



A region may be high in diversity



However, there is an important difference between *the types of cells* within a tissue



Or a region may be high in evenness



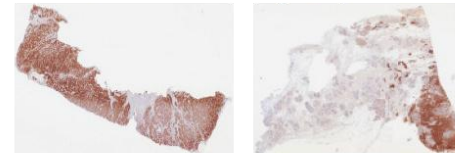
Variability

Tissue	Count	Number of sections sampled in a tissue	Number of tissues	Biological variability across a tissue section (average coefficient of variation)
Variation of different composite regions averaged across a single tissue section				
Human breast	HER2 (+3,+2,+1,0 scoring)	15-20 averaged as one composite region	180	17%
Human breast	ER percent positive cells	15-20 averaged as one composite region	180	11%
Human breast	PR percent positive cells	15-20 averaged as one composite region	180	33%

Table 1: Tissue cross-sections exhibit high levels of biological variability. The table shows typical coefficients of variation by sampling multiple small related sections across a large tissue section using a pathologist. CV values depend on the marker, and are high even when the companion diagnostic scoring rules are applied.

Heterogeneity

Tumor Level Heterogeneity: Het_{TUMOR}

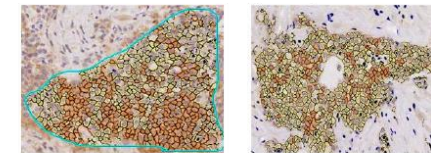


Low tumor heterogeneity

High tumor heterogeneity

Fig. 1: Tumor heterogeneity. Tumor level heterogeneity is the differential expression of a marker within the entire tumor section. The tumor on the left shows very strong HER2 staining, with low heterogeneity across the entire section. The tumor on the right has high HER2 staining in the lower right region, with variation in staining across the remainder of the tumor. Both regions received a summary +3 HER2 score according to ASCO/CAP scoring guidelines, despite clearly having differences in tumor profile.

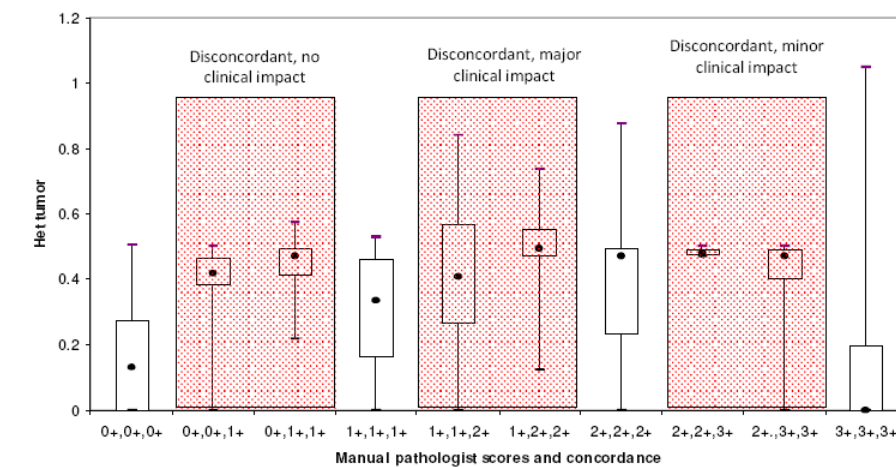
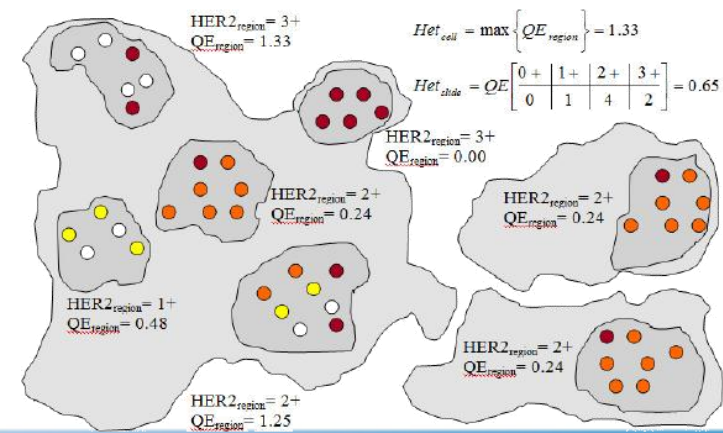
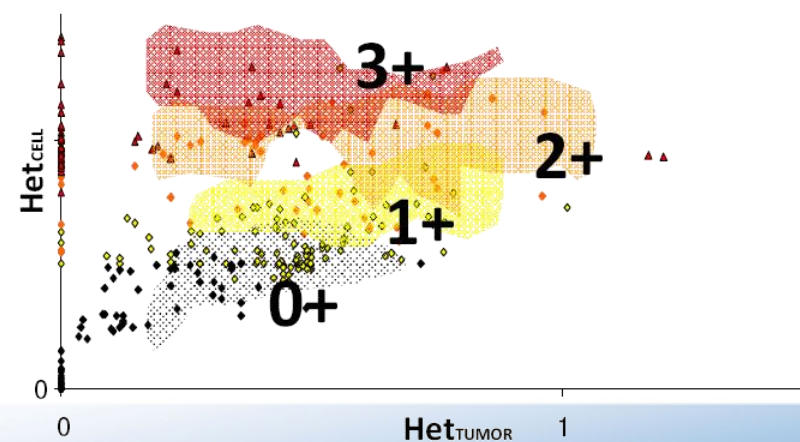
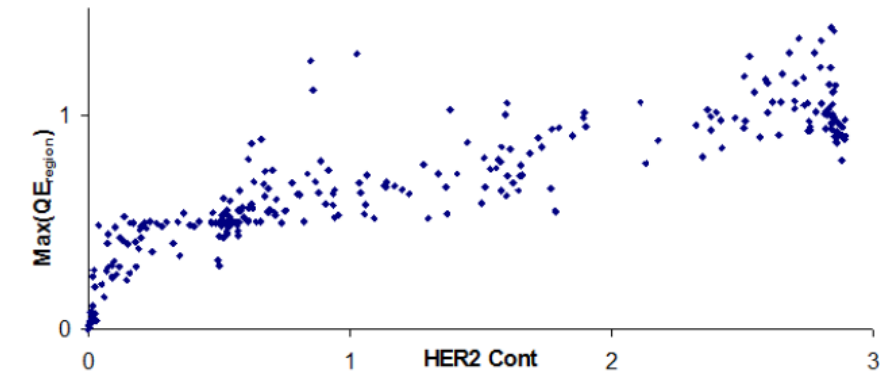
Cell Level Heterogeneity: Het_{CELL}



Low cell heterogeneity

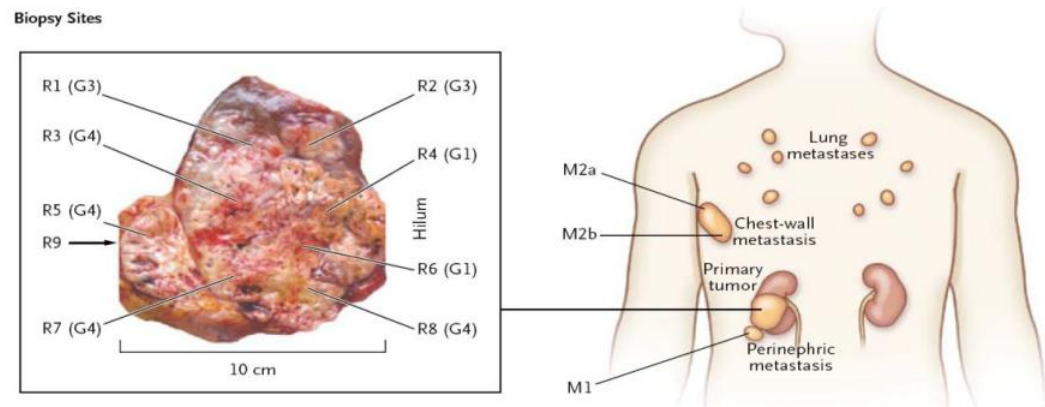
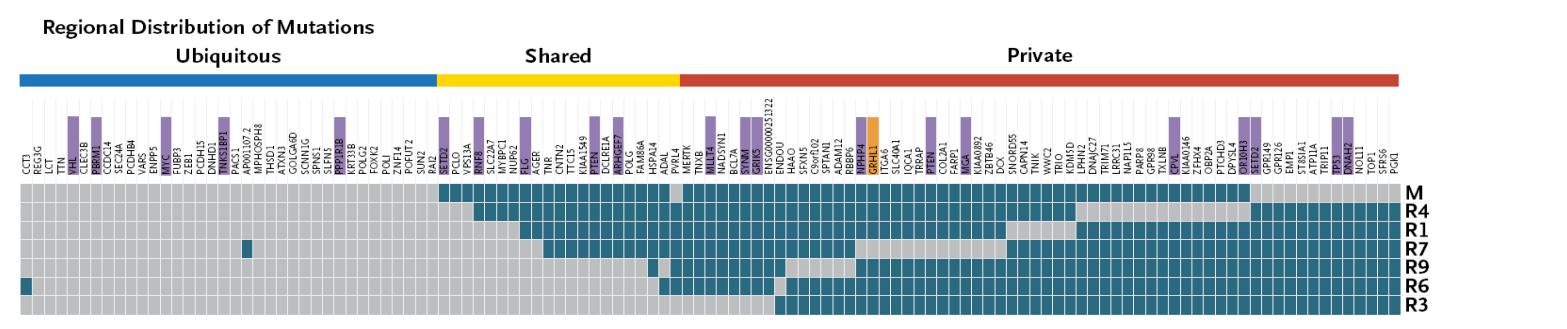
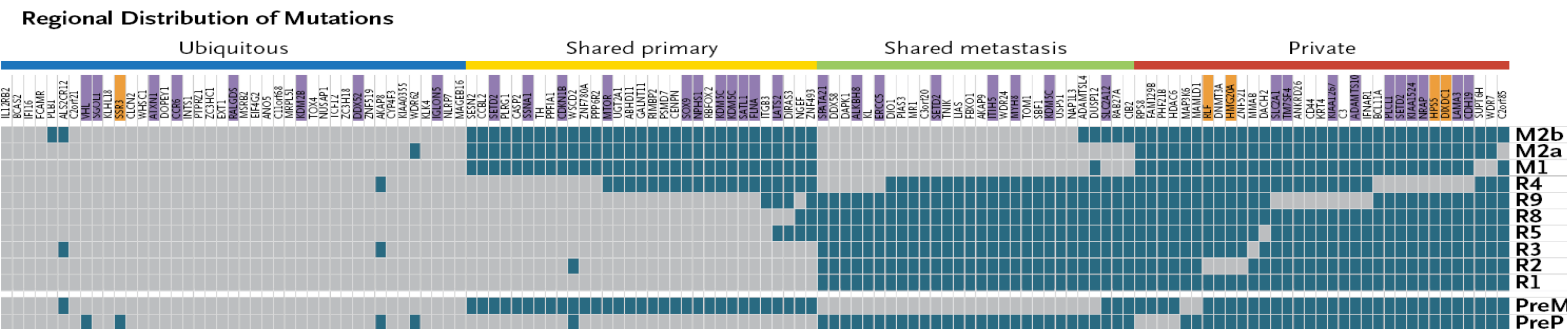
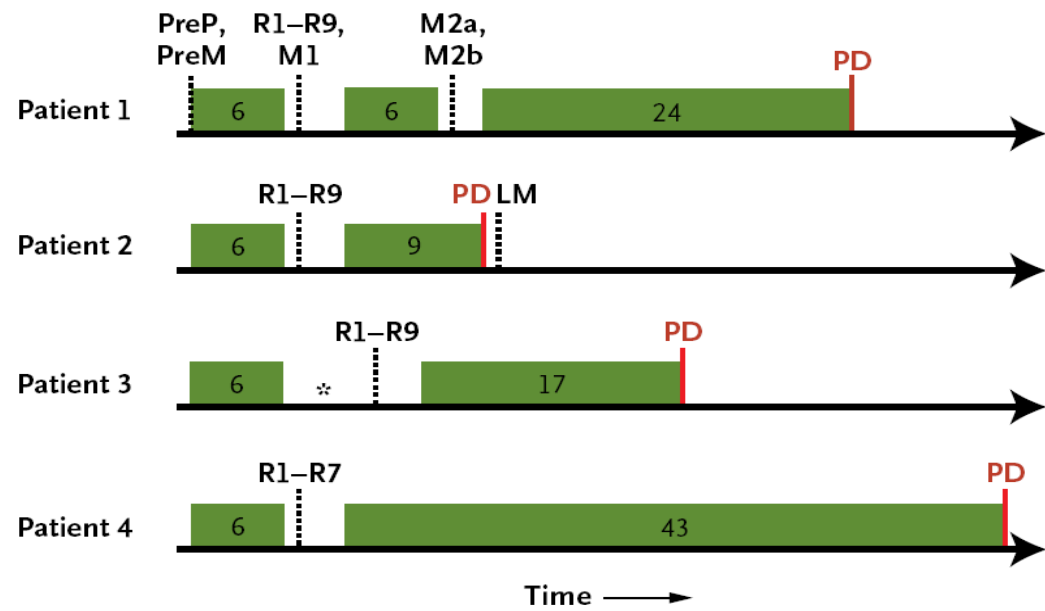
High cell heterogeneity

Figure 2: Cell Heterogeneity. Cell level heterogeneity is the differential expression of a marker from cell-to-cell in a region. The tumor on the left shows low cell-level heterogeneity in the lower right area of the region. The tumor on the right has high cell-level heterogeneity across the region. Both regions received a summary +3 HER2 score according to ASCO/CAP scoring guidelines, despite clearly having different population dynamics.



Intratumor heterogeneity and impact on biomarker detection

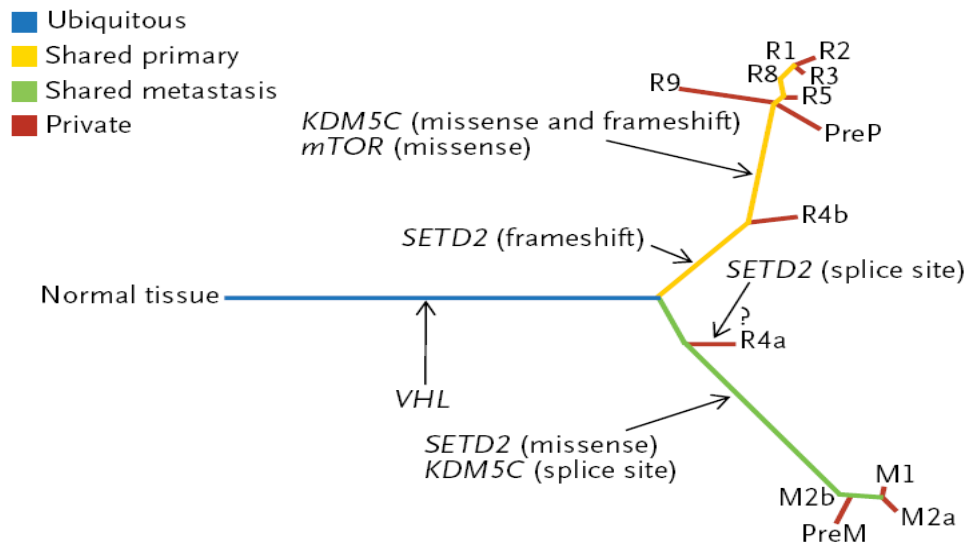
#964 *Intra-tumor heterogeneity and Darwinian selection revealed by multi-region exome sequencing of renal cell carcinomas.* M Gerlinger, A Rowan, S Horswell, J Larkin, D Endesfelder, E Gronroos, P Martinez, N Matthews, A Stewart, P Tarpey, I Varela, B Phillimore, S Begum, N McDonald, A Butler, D Jones, K Raine, C Latimer, C Santos, M Nohadani, A Eklund, B Spencer-Dene, G Clark, L Pickering, G Stamp, M Gore, Z Szallasi, J Downard, PA Futreal, C Swanton



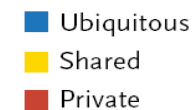
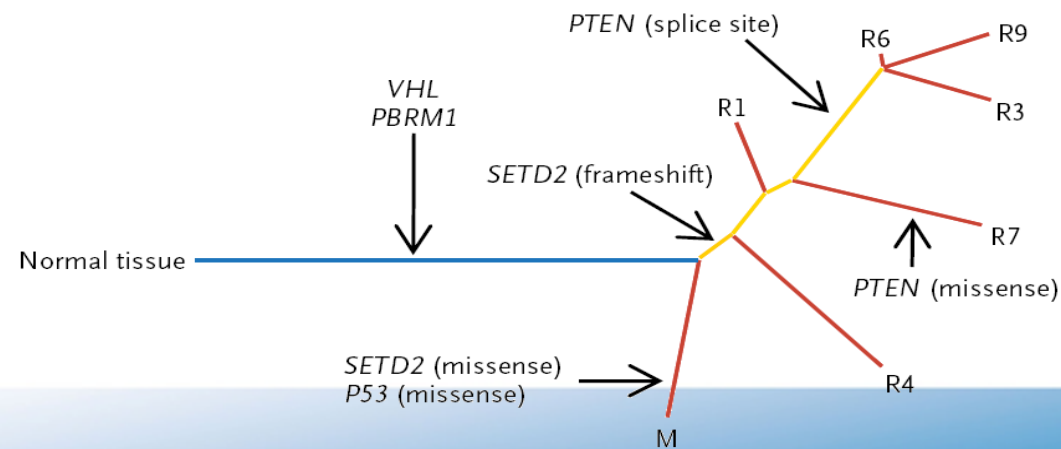
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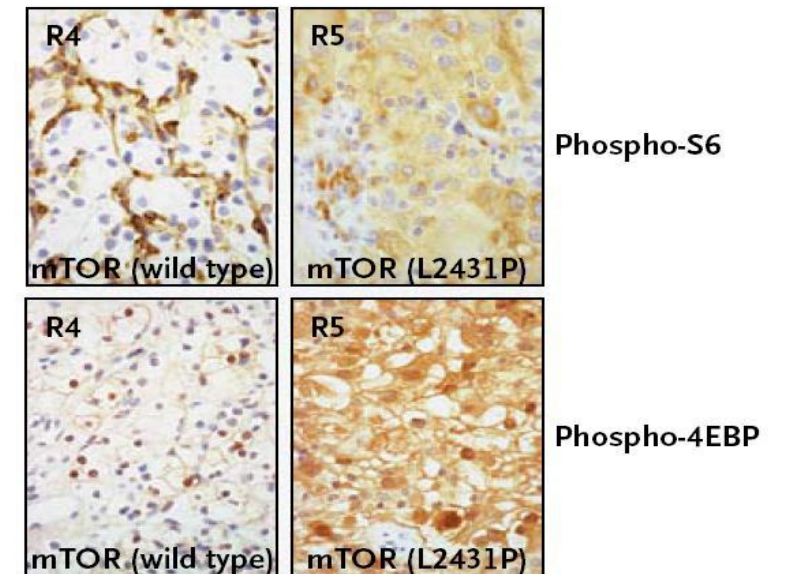
Phylogenetic Relationships of Tumor Regions



Phylogenetic Relationships of Tumor Regions

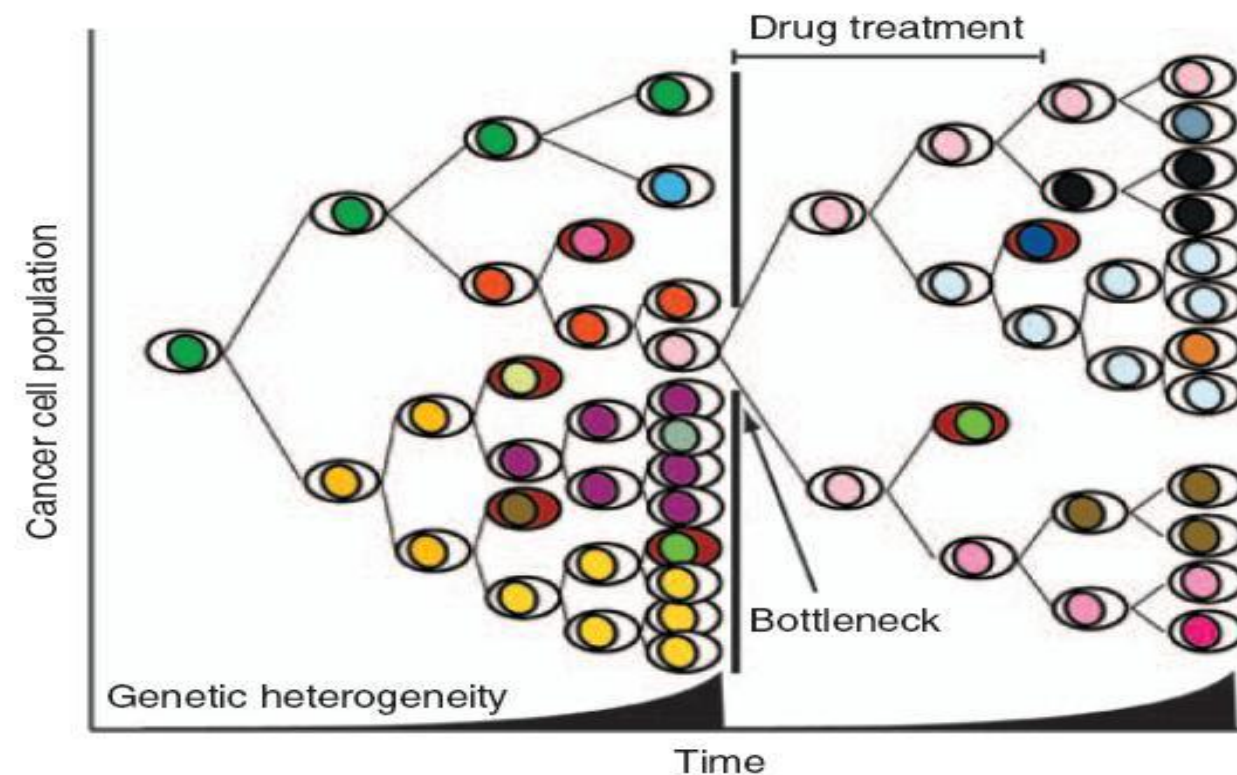


mTOR Staining



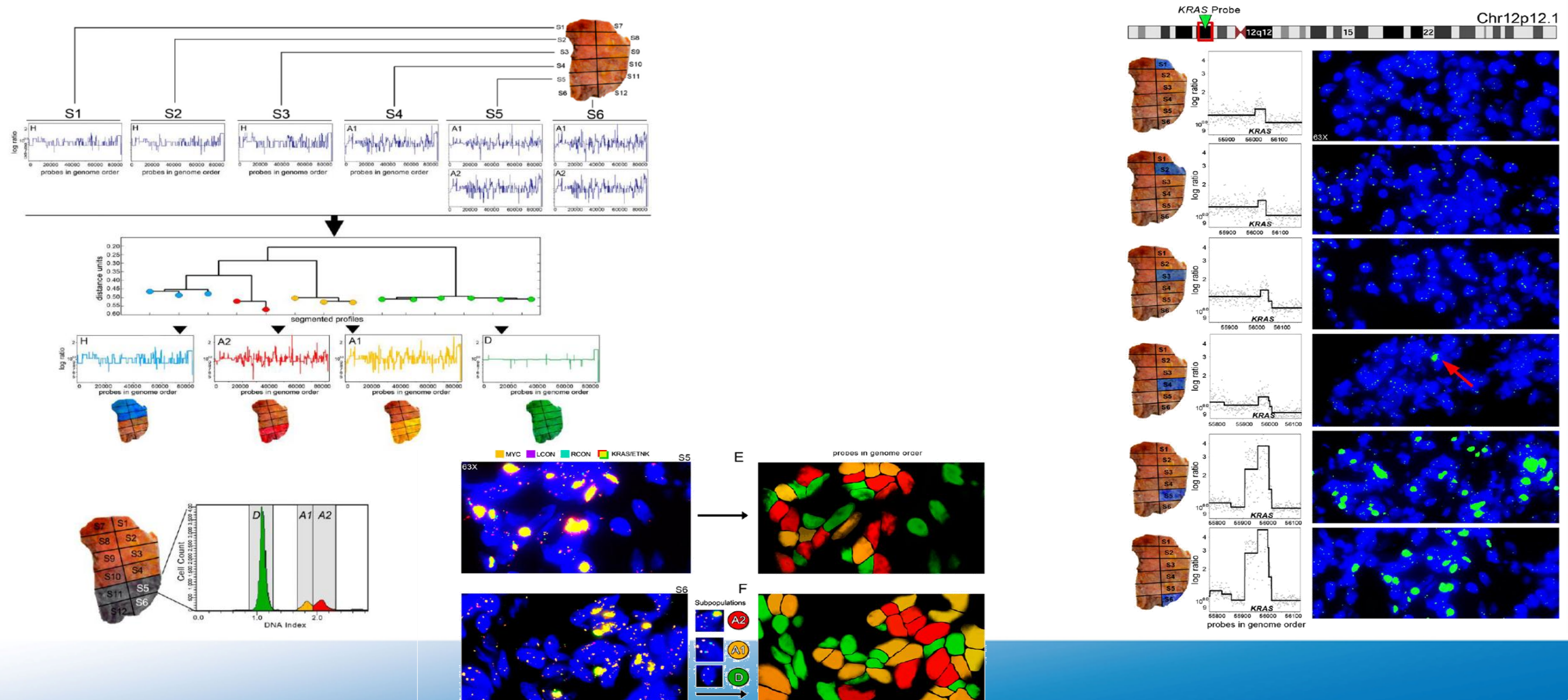
Intratumor heterogeneity and impact on biomarker detection

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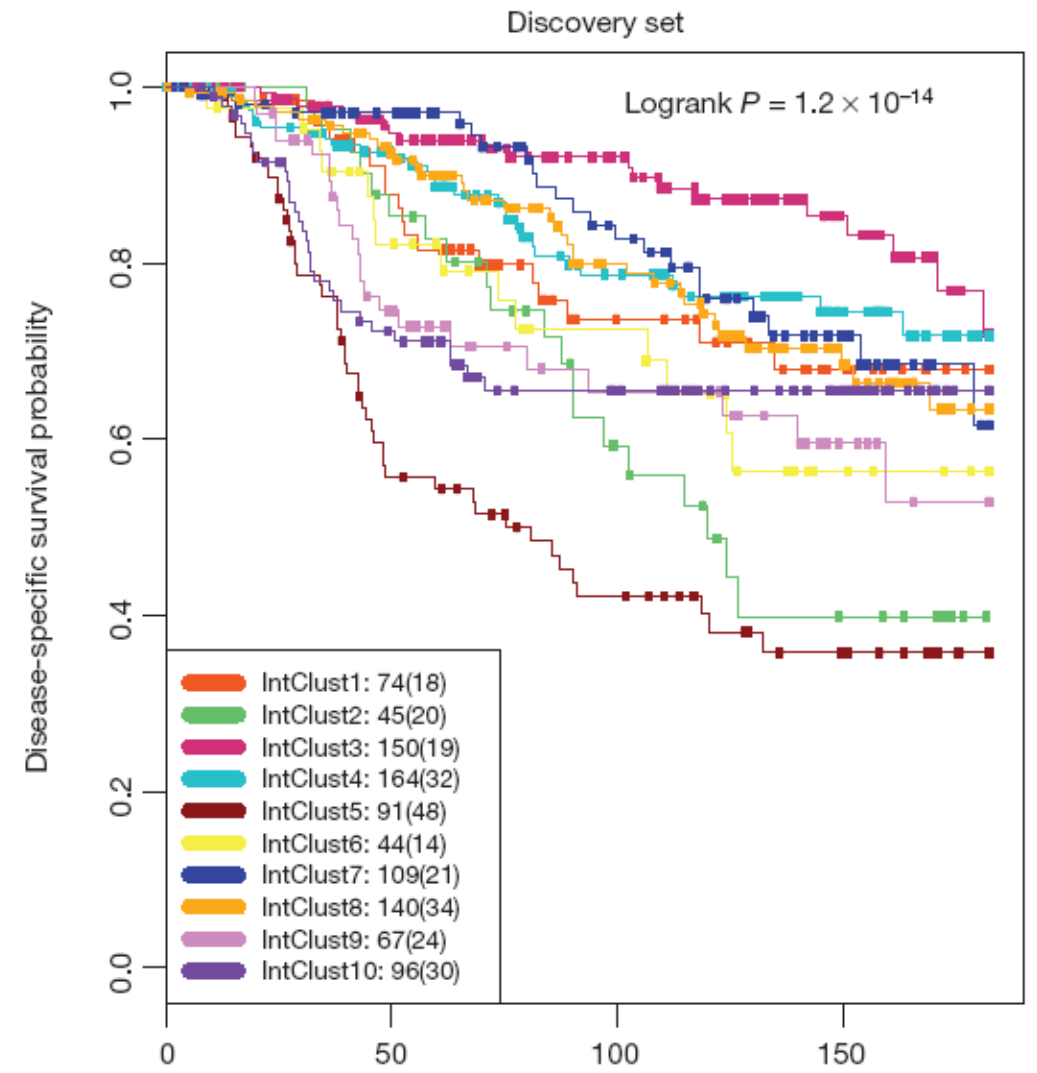
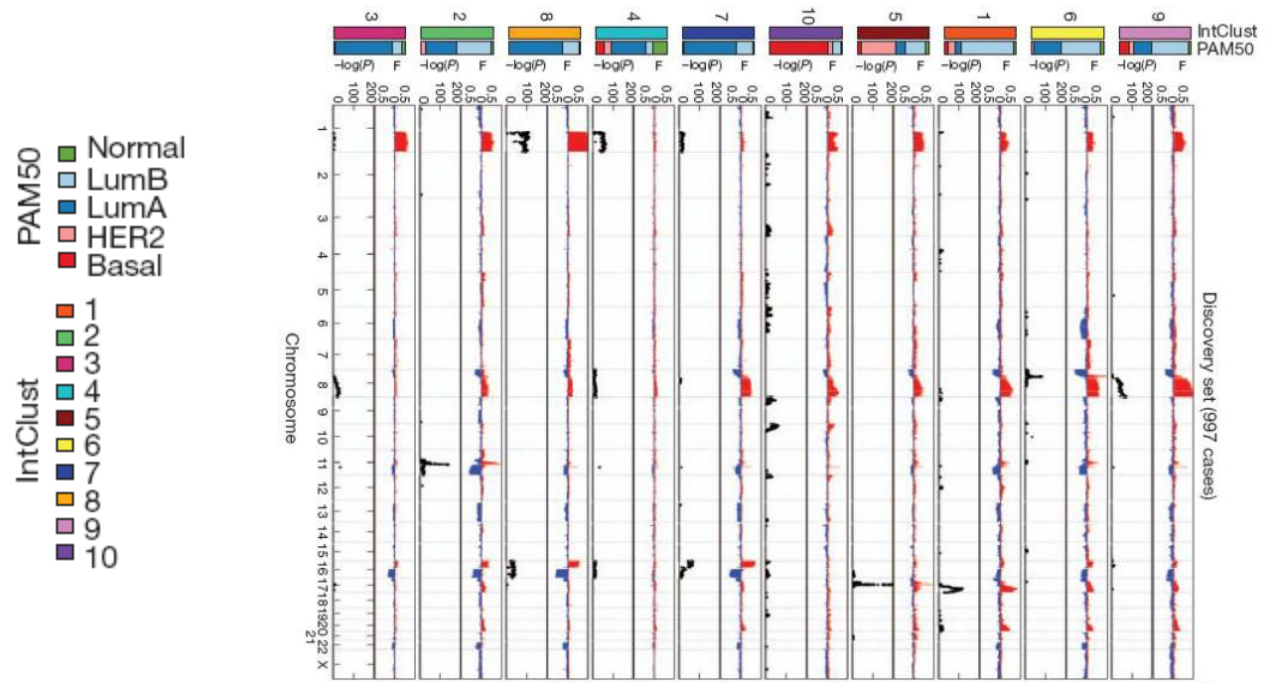
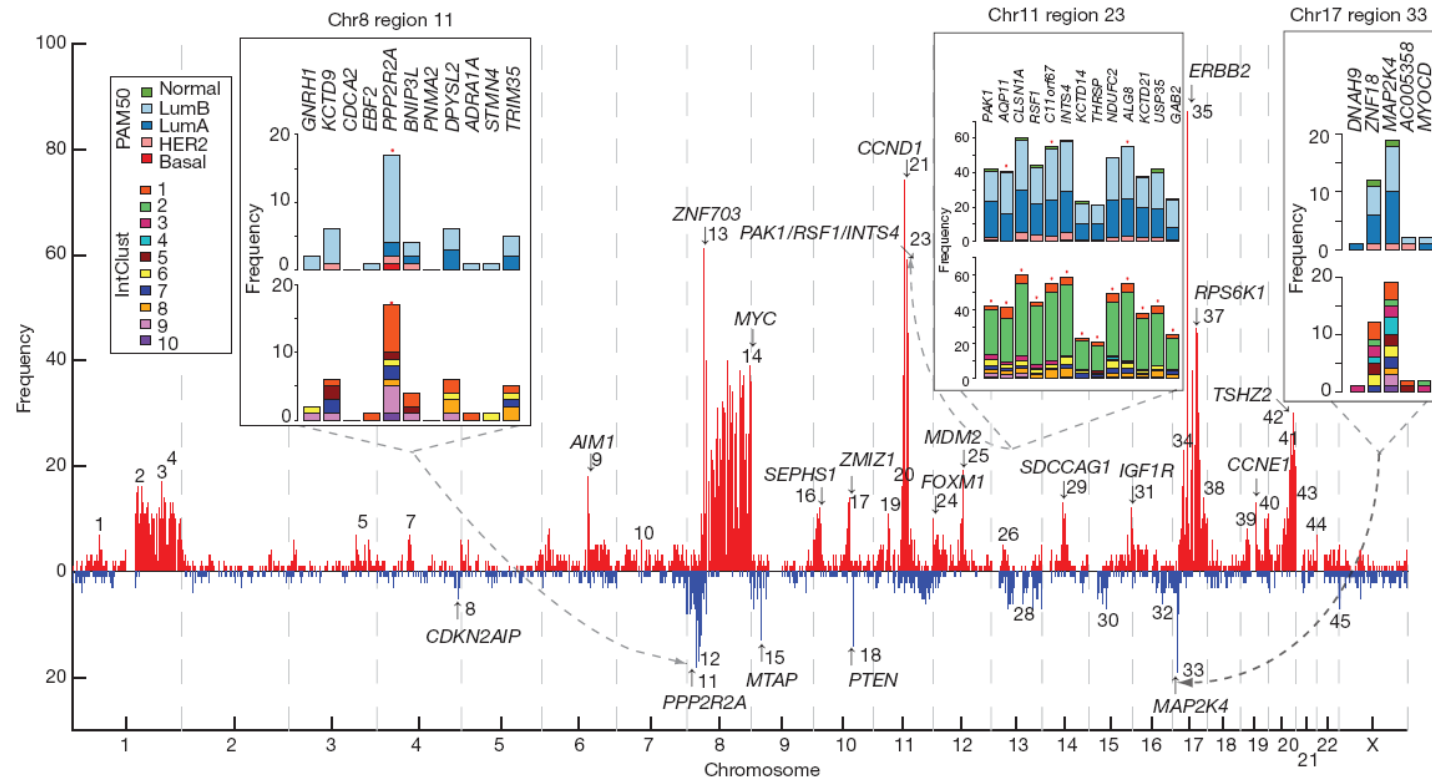
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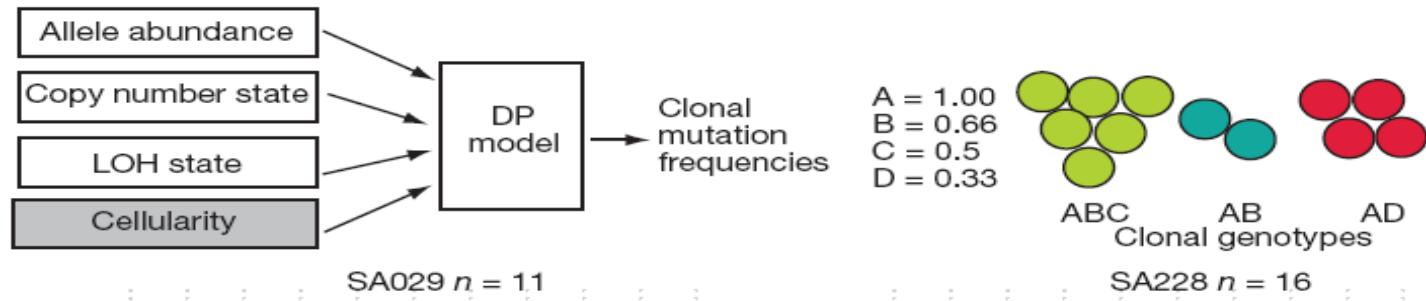
The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups

Christina Curtis^{1,2†*}, Sohrab P. Shah^{3,4*}, Suet-Feung Chin^{1,2*}, Gulisa Turashvili^{3,4*}, Oscar M. Rueda^{1,2}, Mark J. Dunning², Doug Speed^{2,5†}, Andy G. Lynch^{1,2}, Shamith Samarajiwa^{1,2}, Yinyin Yuan^{1,2}, Stefan Gräf^{1,2}, Gavin Ha³, Gholamreza Haffari³, Ali Bashashati³, Roslin Russell², Steven McKinney^{3,4}, METABRIC Group[‡], Anita Langerød⁶, Andrew Green⁷, Elena Provenzano⁸, Gordon Wishart⁸, Sarah Pinder⁹, Peter Watson^{3,4,10}, Florian Markowetz^{1,2}, Leigh Murphy¹⁰, Ian Ellis⁷, Arnie Purushotham^{9,11}, Anne-Lise Børresen-Dale^{6,12}, James D. Brenton^{2,13}, Simon Tavaré^{1,2,5,14}, Carlos Caldas^{1,2,8,13} & Samuel Aparicio^{3,4}



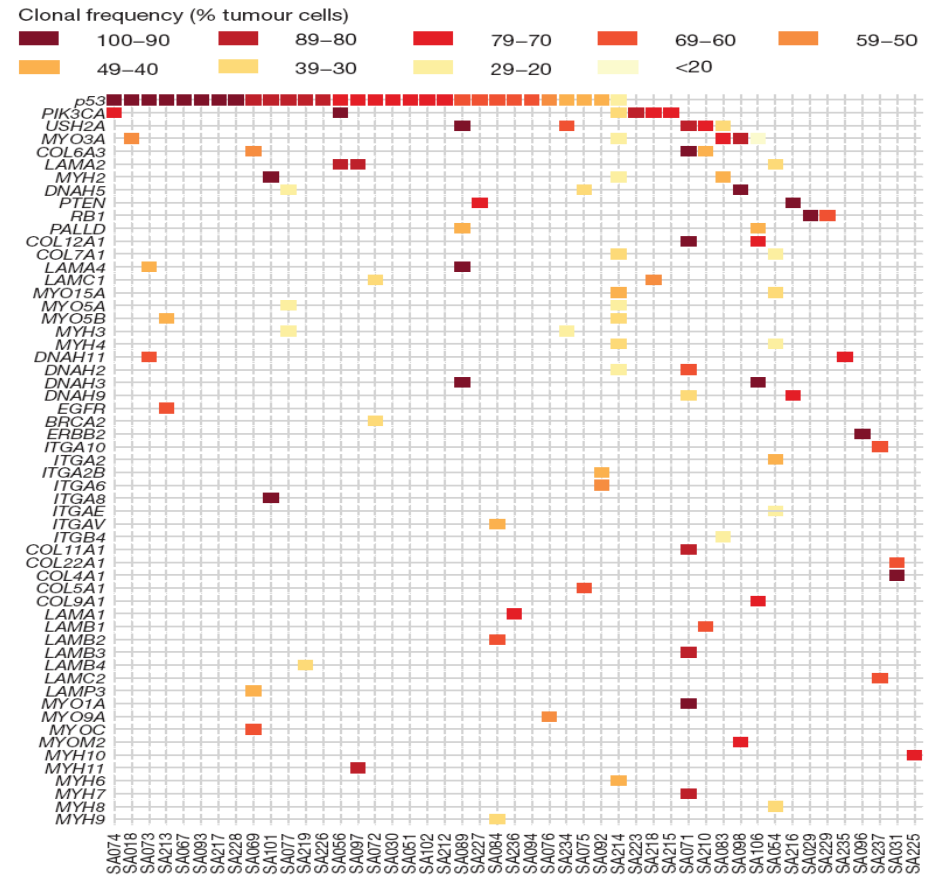
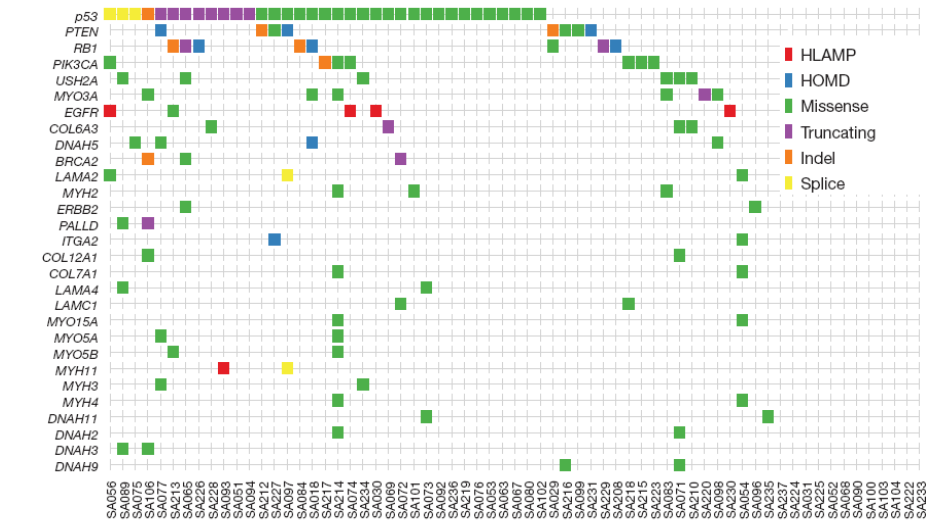
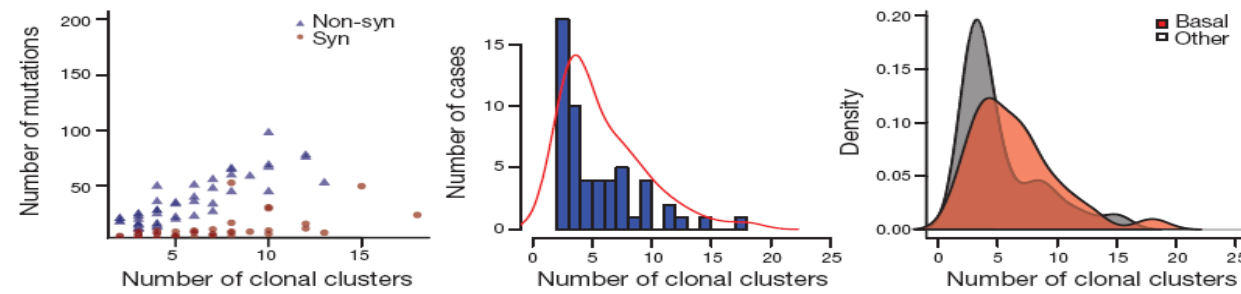
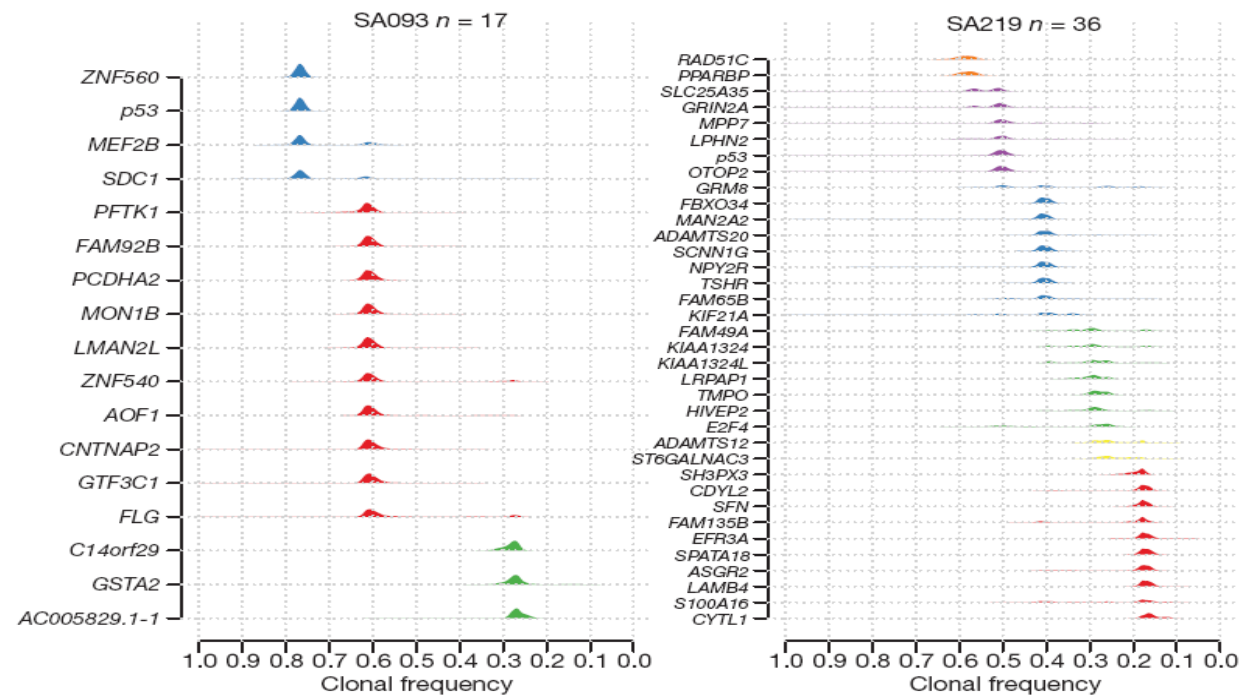
The clonal and mutational evolution spectrum of primary triple-negative breast cancers

Sohrab P. Shah^{1,2}, Andrew Roth^{1,2*}, Rodrigo Goya^{3*}, Arusha Oloumi^{1,2*}, Gavin Ha^{1,2*}, Yongjun Zhao^{3*}, Gulisa Turashvili^{1,2*}, Jiarui Ding^{1,2*}, Kane Tse^{3*}, Gholamreza Haffari^{1,2*}, Ali Bashashati^{1,2*}, Leah M. Prentice^{1,2}, Jaswinder Khattri^{1,2}, Angela Burleigh^{1,2}, Damian Yap^{1,2}, Virginie Bernard⁴, Andrew McPherson^{1,2}, Karey Shumansky^{1,2}, Anamaria Crisan^{1,2}, Ryan Giuliani^{1,2}, Alireza Heravi-Moussavi^{1,2}, Jamie Rosner^{1,2}, Daniel Lai^{1,2}, Inanc Birol³, Richard Varhol³, Angela Tam³, Noreen Dhalla³, Thomas Zeng³, Kevin Ma³, Simon K. Chan³, Malachi Griffith³, Annie Moradian³, S.-W. Grace Cheng³, Gregg B. Morin^{3,5}, Peter Watson^{1,6}, Karen Gelmon⁶, Stephen Chia⁶, Suet-Feung Chin^{7,8}, Christina Curtis^{7,8,9}, Oscar M. Rueda^{7,8}, Paul D. Pharoah⁷, Sambasivarao Damaraju¹⁰, John Mackey¹⁰, Kelly Hoon¹¹, Timothy Harkins¹¹, Vasisht Tadigotla¹¹, Mahvash Sigaroudinia¹², Philippe Gascard¹², Thea Tlsty¹², Joseph F. Costello¹³, Irmtraud M. Meyer^{5,14,15}, Connie J. Eaves¹⁶, Wyeth W. Wasserman^{4,5}, Steven Jones^{3,5,17}, David Huntsman^{1,2,18}, Martin Hirst^{3,15,19}, Carlos Caldas^{7,8,20,21}, Marco A. Marra^{3,5} & Samuel Aparicio^{1,2}



SA029 $n = 11$

SA228 $n = 16$



Mensajes finales

- Emergen nuevos potenciales marcadores de sensibilidad a inhibidores de BRAF en melanoma
- La resistencia a los inhibidores de BRAF en cáncer colorectal se revierte combinando inhibidores de EGFR
- PI3K y PTEN, posibles marcadores de sensibilidad a los inhibidores de mTOR
- Importancia de las mutaciones/variantes de HER2 en la resistencia a los inhibidores de HER2
- Relevancia de la heterogeneidad tumoral